

artus[®] HI Virus-1 QS-RGQ Kit Handbook



24 (catalog no. 4513363)



72 (catalog no. 4513366)

Version 1

IVD

Quantitative in vitro diagnostics

For use with QIA Symphony[®] SP/AS and Rotor-Gene[®] Q Instruments



REF

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QIAGEN GmbH, QIAGEN Strasse 1, 40724 Hilden, GERMANY

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Contents

Intended Use	4
Summary and Explanation	4
Pathogen information	5
Materials Provided	6
Kit contents	6
Materials Required but Not Provided	7
Warnings and Precautions	7
General precautions	8
Reagent Storage and Handling	8
Specimen Handling and Storage	8
Procedure	9
Getting started on the QIA Symphony SP/AS instruments	9
Viral RNA purification	9
Using an internal control and carrier RNA (CARRIER)	9
Assay Control Sets and Assay Parameter Sets	9
Yields of nucleic acids	10
Storing nucleic acids	10
Protocols	
■ RNA isolation and assay setup on the QIA Symphony SP/AS	11
■ RT-PCR on the Rotor-Gene Q	15
Interpretation of Results	16
Troubleshooting guide	16
Quality Control	20
Limitations	20
Performance Characteristics	20
References	21
Symbols	21
Contact Information	22
Ordering Information	23

Intended Use

The *artus* HI Virus-1 QS-RGQ Kit is an in vitro nucleic acid amplification test for the quantitation of human immunodeficiency virus type 1 (HIV-1) RNA in human biological samples. This diagnostic test kit utilizes the reverse transcription polymerase chain reaction (RT-PCR) and is configured for use with the QIASymphony SP/AS and Rotor-Gene Q instruments. Sample containing Group M Subtypes A–H have been validated for use in the assay.

The *artus* HI Virus-1 QS-RGQ Kit is intended for use in conjunction with clinical presentation and other laboratory markers for disease prognosis and for use as an aid in assessing viral response to antiretroviral treatment as measured by changes in human EDTA plasma HIV-1 RNA levels. The *artus* HI Virus-1 QS-RGQ Kit is not intended to be used as a screening test for HIV or as a diagnostic test to confirm the presence of HIV infection.



For more information about specific human biological samples with which the kit has been validated, see the Application Sheets, available online at www.qiagen.com/products/artushivirusrt-pcrkitce.aspx.

As QIAGEN continuously monitors the assay's performance and validates new claims, the users are required to ensure that they work with the latest revision of the instruction for use.



Check availability of new electronic labeling revisions at www.qiagen.com/products/artushivirusrt-pcrkitce.aspx before test execution.

All kits can be used with the respective instruction elements as long as the version number of the handbook and other labeling information matches with the kit version number. The version number is visible on each kit box label. QIAGEN ensures compatibility between all test kit lots under the same version number.

Summary and Explanation

The *artus* HI Virus-1 QS-RGQ Kit constitutes a ready-to-use system for the detection of HIV-1 RNA using polymerase chain reaction (PCR) on Rotor-Gene Q Instruments with sample preparation and assay setup using the QIASymphony SP/AS instruments. The HI Virus-1 RG Master A and B contain reagents and enzymes for the reverse transcription and specific amplification of a 93 bp region of the HIV-1 genome, and for the direct detection of the specific amplicon in fluorescence channel Cycling Green of the Rotor-Gene Q.

In addition, the *artus* HI Virus-1 QS-RGQ Kit contains a second heterologous amplification system to identify possible PCR inhibition. This is detected as an internal control (IC) in fluorescence channel Cycling Orange of the Rotor-Gene Q. The detection limit of the analytical HI Virus-1 RT-PCR is not reduced. External positive controls (HI Virus-1 RG QS 1–4) are supplied, which

allow the determination of the amount of viral RNA. For further information, see the relevant Application Sheet at www.qiagen.com/products/artushivirusrt-pcrkitce.aspx.

Pathogen information

The human immunodeficiency virus (HIV) is a retrovirus that causes acquired immunodeficiency syndrome (AIDS). There are two types of HIV responsible for human infections, HIV-1 and HIV-2, which differ in their virulence and prevalence. Most reported cases of AIDS around the world have been attributed to HIV-1. Infection with HIV occurs by the transfer of infected blood, vaginal fluid, breast milk and other body fluids. Within these body fluids HIV is present as both free virus particles and virus within infected immune cells. The three major routes of transmission are unprotected sexual intercourse, contaminated needles, and transmission from an infected mother to her baby at birth or through breast milk.

HIV primarily infects cells in the human immune system such as helper T cells (specifically CD4⁺). HIV infection leads to low levels of CD4⁺ T cells. When CD4⁺ T cell number decline below a critical level, cell-mediated immunity is lost, and the body becomes progressively more susceptible to opportunistic infections.

AIDS symptoms occur at an advanced stage of HIV infection when the compromised immune system cannot fight off opportunistic infections. At this stage, the infected person increasingly develops symptoms triggered by such infections. The most common infections include chronic cryptosporidium diarrhea, cytomegalovirus-induced eye infection, pneumocystis pneumonia, toxoplasmosis, and tuberculosis as well as infections with members of the *Mycobacterium avium* complex. In addition, the development of different types of cancer, such as invasive cervical cancer, Kaposi sarcoma, or lymphoma, is frequently observed. At present, there is no cure for AIDS, and it is believed that most HIV infected people will eventually die of an AIDS-related illness. However, advancements in HIV/AIDS therapies, including those that fight the virus itself as well as those that prevent or treat opportunistic infections, have drastically improved life expectancy and quality of many HIV/AIDS patients. (1–4)

Materials Provided

Kit contents

artus HI Virus-1 QS-RGQ Kit		(24)	(72)
Catalog no.		4513363	4513366
Number of reactions		24	72
Blue	HI Virus-1 RG Master A	4 x 144 μ l	8 x 144 μ l
Violet	HI Virus-1 RG Master B	4 x 216 μ l	8 x 216 μ l
Red	HI Virus-1 RG QS 1* (1x 10 ⁴ IU/ μ l) QS	200 μ l	200 μ l
Red	HI Virus-1 RG QS 2* (1x 10 ³ IU/ μ l) QS	200 μ l	200 μ l
Red	HI Virus-1 RG QS 3* (1x 10 ² IU/ μ l) QS	200 μ l	200 μ l
Red	HI Virus-1 RG QS 4* (1x 10 ¹ IU/ μ l) QS	200 μ l	200 μ l
Green	HI Virus-1 RG IC [†] IC	1000 μ l	2 x 1000 μ l
White	Water (PCR grade)	1000 μ l	1000 μ l
	<i>artus HI Virus-1 QS-RGQ Kit Handbook</i> (English)	1	1

* Quantitation standard.

† Internal control.

Materials Required but Not Provided

When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, consult the appropriate safety data sheets (SDSs), available from the product supplier.

- Pipets (adjustable)* and sterile pipet tips with filters
- Vortex mixer*
- Benchtop centrifuge* with rotor for 2 ml reaction tubes, capable of centrifugation at 6800 x g

For sample preparation

- QIAasymphony SP instrument (cat. no. 9001297)*
- QIAasymphony AS instrument (cat. no. 9001301)*

For PCR

- Rotor-Gene Q MDx 5plex HRM*†
- Rotor-Gene Q software version 2.1, or higher
- Optional: Rotor-Gene AssayManager version 1.0, or higher

Note: Additional information about materials required for specific applications is contained in the relevant Application Sheet at www.qiagen.com/products/artushivirusrt-pcrkitce.aspx.

Warnings and Precautions

For in vitro diagnostic use

When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, please consult the appropriate safety data sheets (SDSs). These are available online in convenient and compact PDF format at www.qiagen.com/safety where you can find, view, and print the SDS for each QIAGEN® kit and kit component.

For safety information for the purification kit used, see the relevant kit handbook. For safety information regarding instruments, see the relevant instrument user manual.

Discard sample and assay waste according to your local safety regulations.

* Ensure that instruments have been checked and calibrated according to the manufacturer's recommendations.

†If applicable, Rotor-Gene Q 5plex HRM instrument with a production date of January 2010 or later. The production date can be obtained from the serial number on the back of the instrument. The serial number is in the format "mmyynnn" where "mm" indicates the production month in digits, "yy" indicates the last two digits of the production year, and "nnn" indicates the unique instrument identifier.

General precautions

Always pay attention to the following:

- Use sterile pipet tips with filters.
- During manual steps, keep tubes closed when possible and avoid contamination.
- Thaw all components thoroughly at room temperature (15–25°C) before starting an assay.
- When thawed, mix the components (by pipetting repeatedly up and down or by pulse vortexing) and centrifuge briefly. Ensure that no foam or bubbles are present in the reagent tubes.
- Do not mix components from kits with different lot numbers.
- Make sure that the required adapters are precooled to 2–8°C.
- Work quickly and keep PCR reagents on ice or in the cooling block before loading.
- Proceed continuously from one part of the workflow to the next. Do not exceed 30 minutes of transfer time between each module (QIAasymphony SP to QIAasymphony AS to Rotor-Gene Q).

Reagent Storage and Handling

The components of the *artus* HI Virus-1 QS-RGQ Kit should be stored at –15 to –30°C and are stable until the expiration date stated on the label. Repeated thawing and freezing (>2 x) should be avoided, as this may reduce assay performance.

Specimen Handling and Storage

Information about specimen handling and storage for specific applications is contained in the relevant Application Sheet at www.qiagen.com/products/artushivirusrt-pcrkitce.aspx.

Procedure

Getting started on the QIA Symphony SP/AS instruments

Close all drawers and the hoods.

Switch on the QIA Symphony SP/AS instruments, and wait until the “Sample Preparation” screen appears and the initialization procedure has finished.

Log into the instrument (drawers will unlock).

Viral RNA purification

The *artus* HI Virus-1 QS-RGQ Kit has been validated with a viral RNA purification step performed on the QIA Symphony SP using a QIA Symphony DSP Virus/Pathogen Kit. See the *QIA Symphony DSP Virus/Pathogen Handbook* for all the information on how to prepare the reagent cartridge for the sample purification step on the QIA Symphony SP.

Using an internal control and carrier RNA (CARRIER)

Using QIA Symphony DSP Virus/Pathogen Kits in combination with the *artus* HI Virus-1 QS-RGQ Kit requires introduction of the internal control (HI Virus-1 RG IC) into the purification procedure to monitor the efficiency of sample preparation and downstream assay. In addition, QIA Symphony DSP Virus/Pathogen Kits may require the preparation of carrier RNA (CARRIER). For specific information regarding the internal control and the use of carrier RNA (CARRIER), see the relevant Application Sheet at www.qiagen.com/products/artushivirusrtpcrkitce.aspx.

Assay Control Sets and Assay Parameter Sets

Assay Control Sets are the combination of a protocol plus additional parameters, such as internal control, for sample purification on the QIA Symphony SP. A default Assay Control Set is preinstalled for each protocol.

Assay Parameter Sets are the combination of an assay definition with additional parameters defined, such as replicate count and number of assay standards, for assay setup on the QIA Symphony AS.

For integrated runs on the QIA Symphony SP/AS, the Assay Parameter Set is directly linked to an upfront Assay Control Set specifying the associated sample purification process.

Yields of nucleic acids

Eluates prepared with carrier RNA (CARRIER) may contain much more carrier RNA (CARRIER) than target nucleic acids. We recommend using quantitative amplification methods to determine yields.

Storing nucleic acids

For short-term storage of up to 24 hours, we recommend storing purified nucleic acids at 2–8°C. For long-term storage of over 24 hours, we recommend storage at –20°C.

Protocol: RNA isolation and assay setup on the QIA Symphony SP/AS

The following description is a general protocol for using QIA Symphony DSP Virus/Pathogen Kits. Detailed information for a specific application, including volumes and tubes, is provided in the relevant Application Sheet at www.qiagen.com/products/artushivirusrp-pcrkitce.aspx.

Important points before starting

- Ensure that you are familiar with operating the QIA Symphony SP/AS instruments. Refer to the user manuals supplied with your instruments and the most current versions available online at www.qiagen.com/products/qiasymphonymrgg.aspx for operating instructions.
- Before using a reagent cartridge (RC) for the first time, check that Buffers QSL2 and QSB1 in the cartridge (RC) do not contain a precipitate. If necessary, remove the troughs containing Buffers QSL2 and QSB1 from the reagent cartridge (RC) and incubate for 30 minutes at 37°C with occasional shaking to dissolve precipitate. Make sure to replace the troughs in the correct positions. If the reagent cartridge (RC) is already pierced, make sure that the troughs are sealed with Reuse Seal Strips and incubate the complete reagent cartridge (RC) for 30 minutes at 37°C with occasional shaking in a water bath.*
- Try to avoid vigorous shaking of the reagent cartridge (RC) otherwise foam may be generated, which can lead to liquid-level detection problems.
- Work quickly and keep PCR reagents on ice or in the cooling block before loading.
- The reagent volumes are optimized for 24 or 72 reactions per kit per run (cat. nos. 4513363 and 4513366, respectively).
- Before each use, all reagents need to be thawed completely, mixed (by repeated up and down pipetting or by quick vortexing), and centrifuged for at least 3 seconds at 6800 x g. Avoid foaming of the reagents.
- Eluates from the sample preparation and all components of the *artus* HI Virus-1 QS-RGQ Kit have been shown to be stable onboard the instrument for at least the normal time required for sample purification for 96 samples and assay setup of 72 assays, including up to 30 minutes transfer time from the QIA Symphony SP to the QIA Symphony AS and up to 30 minutes transfer time from the QIA Symphony AS to the Rotor-Gene Q.

* Ensure that instruments have been checked, maintained, and calibrated regularly according to the manufacturer's instructions.

Things to do before starting

- Prepare all required mixtures. If needed, prepare mixtures containing carrier RNA (CARRIER) and internal controls just before starting. For more information, see the relevant Application Sheet at www.qiagen.com/products/artushivirusrt-pcrkitce.aspx.
- Before starting the procedure, ensure that the magnetic particles are fully resuspended. Vortex the trough containing the magnetic particles vigorously for at least 3 minutes before first use.
- Before loading the reagent cartridge (RC), remove the cover from the trough containing the magnetic particles and open the enzyme tubes. Make sure that the enzyme rack has been equilibrated to room temperature (15–25°C).
- Make sure that the piercing lid (PL) is placed on the reagent cartridge (RC) and the lid of the magnetic-particle trough has been removed or, if using a partially used reagent cartridge (RC), make sure the Reuse Seal Strips have been removed.
- If samples are bar coded, orient samples in the tube carrier so that the bar codes face the bar code reader within the “Sample” drawer at the left side of the QIA Symphony SP.

Procedure

Viral RNA purification on the QIA Symphony SP

1. **Close all drawers and the hoods of the QIA Symphony SP/AS instruments.**
2. **Switch on the instruments, and wait until the “Sample Preparation” screen appears and the initialization procedure has finished.**
The power switch is located at the bottom, left corner of the QIA Symphony SP.
3. **Log in to the instruments.**
4. **Prepare the following drawers according to the relevant Application Sheet at www.qiagen.com/products/artushivirusrt-pcrkitce.aspx.**
 - “Waste” drawer; when prepared, perform an inventory scan.
 - “Eluate” drawer; when prepared, perform an inventory scan.
 - “Reagents and Consumables” drawer; when prepared, perform an inventory scan.
 - “Sample” drawer

5. **Using the “Integrated run” setup on the QIA Symphony touchscreen, enter the required information for each batch of samples to be processed. Select an Assay Parameter Set for the run, and assign it and the corresponding AS batch to the samples.**

Information about the Assay Parameter Set and preselected elution volume is provided on the relevant Application Sheet.

For more information about integrated runs on the QIA Symphony SP/AS, see the instrument user manuals.

6. **When setting up an integrated run, check for correct assignment of sample labware, sample type (sample, EC+, and EC–), and volumes.**

Information about consumables and components to load in each drawer is provided on the relevant Application Sheet.

7. **After information about all batches of the integrated run has been entered, click the “Ok” button to exit the “Integrated run” setup. The status of all batches within the overview of the integrated run changes from “LOADED” to “QUEUED”. As soon as one batch is queued the “Run” button appears. Press the “Run” button to start the procedure.**

All processing steps are fully automated.

Loading the QIA Symphony AS drawers for assay setup

8. **After queuing an integrated run, open the QIA Symphony AS drawers. The required components to be loaded are shown on the touchscreen.**

9. **Always make sure to do the following before the integrated run.**

- Insert the tip chute.
- Discard the tip disposal bag
- Install an empty tip disposal bag

10. **Define and load assay rack(s). Assay rack(s), in precooled adapter(s), are loaded onto the “Assay” slot(s). Information about the assay racks is provided on the relevant Application Sheet at www.qiagen.com/products/artushivirusrt-pcrkitce.aspx.**

11. **Check the temperature of the cooling positions.**

When the target cooling temperatures are reached, the small asterisk next to each slot will appear green.

12. **Combine all tubes of HI Virus-1 RG Master A in a single kit into one tube before use. Combine all tubes of HI Virus-1 RG Master B in a single kit into one tube before use.**

Note: Viscous reagents can be difficult to handle with manual pipets. Make sure to transfer the entire volume of the Master in the tube.

- 13. Fill each reagent tube with the required volume of appropriate reagent according to the loading information given by the instrument software.**

Note: Before each use, all reagents need to be thawed completely, mixed (by repeated up and down pipetting or by quick vortexing), and centrifuged for at least 3 seconds at 6800 x g. Avoid bubbles or foaming, which could cause detection errors. Work quickly and keep PCR components on ice or in the cooling block before loading.

- 14. Load the reagent rack, and place the reagent tubes, without lids, into the appropriate positions of precooled adapters for reagents according to the relevant Application Sheet.**
- 15. Load disposable filter-tips into the “Eluate and Reagents” and “Assays” drawers, according to the required number of each tip type indicated on the relevant Application Sheet.**
- 16. Close the “Eluate and Reagents” and “Assays” drawers.**
- 17. Upon closing each drawer, press “Scan” to start the inventory scan for each drawer.**

The inventory scan checks the slots, adapters, filter-tips, and the tip chute, as well as the correct loading of specific reagent volumes. If required, correct any errors.

The assay setup will start automatically after the purification step on the QIASymphony SP is completed and the eluate racks are transferred to the QIASymphony AS.

- 18. After the run is finished, press “Remove” in the assay setup “Overview” screen. Open the “Assays” drawer and unload the assay rack(s).**
- 19. Download the result and cyclor files.**
- 20. If multiple batches on the QIASymphony AS are configured in an integrated run, reload the QIASymphony AS drawers, starting at step 8.**
- 21. Proceed to “Protocol: RT-PCR on the Rotor-Gene Q”, page 15.**
- 22. Perform the regular maintenance of the QIASymphony AS during the PCR run on the Rotor-Gene Q or later.**

Since the workflow is an integrated operation, clean all instruments at the end of the completed workflow.

Follow the maintenance instructions in the *QIASymphony SP/AS User Manual — General Description*. Make sure to carry out maintenance regularly to minimize the risk of cross-contamination.

Protocol: RT-PCR on the Rotor-Gene Q

Important points before starting

- Take time to familiarize yourself with the Rotor-Gene Q before starting the protocol. See the instrument user manual.
- For automatic interpretation of the PCR results, Rotor-Gene AssayManager may be used instead of Rotor-Gene Q software.
- Make sure that at all 4 quantitation standards as well as at least one negative control (Water, PCR grade) are included per PCR run. To generate a standard curve, use all 4 quantitation standards supplied (HI Virus-1 QS 1–4) for each PCR run.

Procedure

1. **Close the PCR tubes, and place them in the 72-Well Rotor of the Rotor-Gene Q. Make sure to transfer the Rotor-Gene Q 4-strip tubes in the correct orientation, so that the position indices of the cooling adapter and the rotor match. Make sure that the locking ring (accessory of the Rotor-Gene Instrument) is placed on top of the rotor to prevent accidental opening of the tubes during the run.**
2. **Transfer the cyclor file from the QIASymphony AS to the Rotor-Gene Q computer.**
3. **For the detection of HIV-1 RNA, create a temperature profile and start the run according to the relevant Application Sheet at www.qiagen.com/products/artushivirusrt-pcrkitce.aspx. Software-specific information about programming the Rotor-Gene Q is provided in the relevant Protocol Sheet “Settings to run *artus* QS-RGQ Kits” at www.qiagen.com/products/artushivirusrt-pcrkitce.aspx.**

Interpretation of Results

See the relevant Application Sheet at www.qiagen.com/products/artushivirusrt-pcrkitce.aspx for detailed information about interpretation of results.

Troubleshooting guide

This troubleshooting guide may be helpful in solving any problems that may arise. The scientists in QIAGEN Technical Services are always happy to answer any questions you may have about either the information and protocols in this handbook or sample and assay technologies (for contact information, see back cover or visit www.qiagen.com).

Comments and suggestions

General handling

Error message displayed in the touchscreen	If an error message is displayed during a protocol run, refer to the user manuals supplied with your instruments.
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Precipitate in reagent trough of opened cartridge of the QIASymphony DSP Virus/Pathogen Kit

- | | |
|--------------------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| a) Buffer evaporation | Excessive evaporation may lead to increased salt concentration or decreased alcohol concentrations in buffers. Discard reagent cartridge (RC). Make sure to seal buffer troughs of a partially used reagent cartridge (RC) with Reuse Seal Strips when not being used for purification. |
| b) Storage of reagent cartridge (RC) | Storage of reagent cartridge (RC) under 15°C may lead to formation of precipitates. If necessary, remove the troughs containing Buffers QSL2 and QSB1 from the reagent cartridge (RC) and incubate in a water bath* at 37°C for 30 minutes with occasional shaking to dissolve precipitate. Make sure to replace the troughs in the correct positions. If the reagent cartridge (RC) is already pierced, make sure that the troughs are reclosed with Reuse Seal Strips and incubate the complete reagent cartridge (RC) in a water bath* at 37°C for 30 minutes with occasional shaking. |

* Ensure that instruments have been checked, maintained, and calibrated regularly according to the manufacturer's instructions.

Comments and suggestions

Low yield of nucleic acids

- | | |
|---------------------------------------------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| a) Magnetic particles were not completely resuspended | Before starting the procedure, ensure that the magnetic particles are fully resuspended. Vortex for at least 3 minutes before use. |
| b) Frozen samples were not mixed properly after thawing | Thaw frozen samples with mild agitation to ensure thorough mixing. |
| c) Carrier RNA (CARRIER) not added | Reconstitute carrier RNA (CARRIER) in Buffer AVE (AVE) and mix with appropriate volume of Buffer AVE (AVE) as described in the relevant Application Sheet at www.qiagen.com/products/artushivirusrt-pcrkitce.aspx . Repeat the purification procedure with new samples. |
| d) Degraded nucleic acids | Samples were stored incorrectly or subjected to too many freeze–thaw cycles. Repeat the purification procedure with new samples. |
| e) Incomplete sample lysis | Before use, check that Buffer QSL2 and QSB1 do not contain precipitates. If necessary, remove the troughs containing Buffers QSL1 and QSB1 from the reagent cartridge (RC) and incubate for 30 minutes at 37°C with occasional shaking to dissolve precipitate. If the reagent cartridge (RC) is already pierced, make sure that the troughs are reclosed with Reuse Seal Strips, and incubate the complete reagent cartridge (RC) for 30 minutes at 37°C with occasional shaking in a water bath.* |
| f) Clogging of pipet tip due to insoluble material | Insoluble material was not removed from the sample prior to starting the QIA Symphony purification procedure. To remove insoluble material for viral applications, centrifuge the sample at 3000 x g for 1 minute, and transfer the supernatant to a new sample tube. |

*Ensure that instruments have been checked, maintained, and calibrated regularly according to the manufacturer's instructions.

Comments and suggestions

QIA Symphony AS detects insufficient Master

Not all of the Master transferred to tube

Combine all tubes of HI Virus-1 RG Master A in a single kit into one tube before use. Combine all tubes of HI Virus-1 RG Master B in a single kit into one tube before use. Viscous reagents can be difficult to handle with manual pipets. Make sure to transfer the entire volume of the Master in the tube.

For viscous reagents, we recommend aspirating an extra volume of 5% when using manual pipets (e.g., adjust the pipet to 840 μ l for an 800 μ l volume).

Alternatively, after slowly dispensing the liquid and performing a blowout at the target tube's wall, remove the tip from the liquid, release the pipet plunger, and wait for an additional 10 seconds. Residual liquid will flow down the tip and can be blown out by pressing the pipet plunger a second time. The use of PCR grade filter-tips labeled as "low retention" can improve the recovery of liquid.

No signal with positive controls (HI Virus-1 RG QS 1–4) in fluorescence channel Cycling Green

a) The selected fluorescence channel for PCR data analysis does not comply with the protocol

For data analysis select the fluorescence channel Cycling Green for the analytical HI Virus-1 PCR and the fluorescence channel Cycling Orange for the internal control PCR.

b) Incorrect programming of the temperature profile of the Rotor-Gene instrument

Compare the temperature profile with the protocol. See the relevant Application Sheet and Protocol Sheet at www.qiagen.com/products/artushivirusrt-pcrkitce.aspx.

c) Incorrect configuration of the PCR

Make sure that assay setup was performed correctly and that the correct Assay Parameter Set was used. Repeat the PCR, if necessary. See the relevant Application Sheet at www.qiagen.com/products/artushivirusrt-pcrkitce.aspx.

Comments and suggestions

- d) The storage conditions for one or more kit components did not comply with the instructions given in “Reagent Storage and Handling” (page 8) Check the storage conditions and the expiration date (see the kit label) of the reagents and use a new kit, if necessary.
- e) The *artus* HI Virus-1 QS-RGQ Kit has expired Check the storage conditions and the expiration date (see the kit label) of the reagents and use a new kit, if necessary.

Weak or no signal of the internal control of a negative plasma sample subjected to purification using the QIASymphony DSP Virus/Pathogen Kit in fluorescence channel Cycling Orange and simultaneous absence of a signal in channel Cycling Green

- a) The PCR conditions do not comply with the protocol Check the PCR conditions (see above) and repeat the PCR with corrected settings, if necessary.
- b) The PCR was inhibited Make sure that you use the validated isolation method (see “Protocol: RNA isolation and assay setup on the QIASymphony SP/AS”, page 11) and closely follow the instructions.
- c) RNA was lost during extraction An absent signal of the internal control can indicate the loss of RNA during the extraction. Make sure that you use the validated isolation method (see “Protocol: RNA isolation and assay setup on the QIASymphony SP/AS”, page 11) and closely follow the instructions.
See also “Low yield of nucleic acids”, above.
- d) The storage conditions for one or more kit components did not comply with the instructions given in “Reagent Storage and Handling” (page 8) Check the storage conditions and the expiration date (see the kit label) of the reagents and use a new kit, if necessary.
- e) The *artus* HI Virus-1 QS-RGQ Kit has expired Check the storage conditions and the expiration date (see the kit label) of the reagents and use a new kit, if necessary.

Comments and suggestions

Signals with the negative controls in fluorescence channel Cycling Green of the analytical PCR

- | | |
|---------------------------------------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| a) Contamination occurred during preparation of the PCR | Repeat the PCR with new reagents in replicates.
If possible, close the PCR tubes directly after addition of the sample to be tested.
Make sure that work space and instruments are decontaminated at regular intervals. |
| b) Contamination occurred during extraction | Repeat the extraction and PCR of the sample to be tested using new reagents.
Make sure that work space and instruments are decontaminated at regular intervals. |

Quality Control

In accordance with QIAGEN's ISO-certified Quality Management System, each lot of *artus* HI Virus-1 QS-RGQ Kit is tested against predetermined specifications to ensure consistent product quality.

Limitations

All reagents may exclusively be used in in vitro diagnostics.

The product is to be used by personnel specially instructed and trained in the in vitro diagnostics procedures only.

Strict compliance with the user manual is required for optimal PCR results.

Attention should be paid to expiration dates printed on the box and labels of all components. Do not use expired components.

Although rare, mutations within the highly conserved regions of the viral genome covered by the kit's primers and/or probe may result in underquantitation or failure to detect the presence of the virus in these cases. Validity and performance of the assay design are revised at regular intervals.

Performance Characteristics

See www.qiagen.com/products/artushivirusrt-pcrkitce.aspx for performance characteristics of the *artus* HI Virus-1 QS-RGQ Kit.

References

1. McCutchan, F.E. (2006) Global epidemiology of HIV. *J. Med. Virol.* **78** **Suppl 1**, S7.
2. Nikolopoulos, G., Tsiodras, S., Bonovas, S., and Hatzakis, A. (2012) Antiretrovirals for HIV exposure prophylaxis. *Curr. Med. Chem.* **19**, 5924.
3. Perrin, L., Kaiser, L., and Yerly, S. (2003) Travel and the spread of HIV-1 genetic variants. *Lancet Infect. Dis.* **3**, 22.
4. Roques, P. et al. (2004) Phylogenetic characteristics of three new HIV-1 N strains and implications for the origin of group N. *AIDS* **18**, 1371.

Symbols



Contains reagents sufficient for <N> reactions



Use by



In vitro diagnostic medical device



Catalog number



Lot number



Material number



Components



Contains



Number



Global Trade Item Number



Temperature limitation



Manufacturer



Consult instructions for use



Caution

Contact Information

For technical assistance and more information, please see our Technical Support Center at www.qiagen.com/Support, call 00800-22-44-6000, or contact one of the QIAGEN Technical Service Departments or local distributors (see back cover or visit www.qiagen.com).

Ordering Information

Product	Contents	Cat. no.
<i>artus</i> HI Virus-1 QS-RGQ Kit (24)	For 24 reactions: 2 Masters, 4 Quantitation Standards, Internal Control, Water (PCR grade)	4513363
<i>artus</i> HI Virus-1 QS-RGQ Kit (72)	For 72 reactions: 2 Masters, 4 Quantitation Standards, Internal Control, Water (PCR grade)	4513366
QIASymphony RGQ system		
QIASymphony RGQ, System	QIASymphony SP, QIASymphony AS, Rotor-Gene Q MDx 5plex HRM, required accessories and consumables, installation and training	9001850

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www.qiagen.com

Australia ■ techservice-au@qiagen.com

Austria ■ techservice-at@qiagen.com

Belgium ■ techservice-bnl@qiagen.com

Brazil ■ suportetecnico.brasil@qiagen.com

Canada ■ techservice-ca@qiagen.com

China ■ techservice-cn@qiagen.com

Denmark ■ techservice-nordic@qiagen.com

Finland ■ techservice-nordic@qiagen.com

France ■ techservice-fr@qiagen.com

Germany ■ techservice-de@qiagen.com

Hong Kong ■ techservice-hk@qiagen.com

India ■ techservice-india@qiagen.com

Ireland ■ techservice-uk@qiagen.com

Italy ■ techservice-it@qiagen.com

Japan ■ techservice-jp@qiagen.com

Korea (South) ■ techservice-kr@qiagen.com

Luxembourg ■ techservice-bnl@qiagen.com

Mexico ■ techservice-mx@qiagen.com

The Netherlands ■ techservice-bnl@qiagen.com

Norway ■ techservice-nordic@qiagen.com

Singapore ■ techservice-sg@qiagen.com

Sweden ■ techservice-nordic@qiagen.com

Switzerland ■ techservice-ch@qiagen.com

UK ■ techservice-uk@qiagen.com

USA ■ techservice-us@qiagen.com

