DNeasy® Plant Maxi Kit

The DNeasy Plant Maxi Kit (cat. no. 68163) should be stored dry at 2–8°C upon arrival and are stable for up to 1 year under these conditions.

For more information, please refer to the *DNeasy Plant Handbook*, which can be found at www.qiagen.com/handbooks.

For technical assistance, please call toll-free 00800-22-44-6000, or find regional phone numbers at www.qiagen.com/contact.

Notes before starting

- Perform all centrifugation steps at 3000–5000 x *g* (5000 x *g* is preferable) at room temperature (15–25°C) in a centrifuge with a swing-out rotor.
- If necessary, redissolve any precipitates in Buffer AP1 and Buffer AW1 concentrates.
- Add ethanol to Buffer AW1 and Buffer AW2 concentrates.
- Preheat a water bath or heating block to 65°C.
- Preheat Buffer AP1 to 65°C.

1. Disrupt samples (≤1 g wet weight or ≤0.2 g lyophilized tissue) using the TissueRuptor®, the TissueLyser II, or a mortar and pestle.
2. Add 5 ml preheated Buffer AP1 and 10 µl RNase A. Vortex vigorously until no tissue clumps are visible. Incubate for 10 min at 65°C. Invert tube 2–3 times during incubation.

   Note: Do not mix Buffer AP1 and RNase A before use.
3. Add 1.8 ml Buffer P3. Mix and incubate for 10 min on ice.
4. Centrifuge at 3000–5000 x *g* for 5 min.
5. Decant the supernatant into a QIAshredder spin column placed in a 50 ml collection tube. Centrifuge at 3000–5000 x *g* for 5 min.

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6. Transfer the flow-through, without disturbing the pellet if present, into a new 50 ml tube. Add 1.5 volumes Buffer AW1, and mix by vortexing.

   Note: It is important to add Buffer AW1 to the lysate and to mix immediately.

7. Transfer the mixture from step 6 (maximum 15 ml) into a DNeasy Maxi spin column placed in a 50 ml collection tube. Centrifuge at 3000–5000 x *g* for 5 min. Discard the flow-through.

8. Add 12 ml Buffer AW2, and centrifuge for 10 min at 3000–5000 x *g*. Discard the flow-through.

9. Transfer the spin column to a new 50 ml tube.

10. Add 0.75–1.0 ml Buffer AE for elution. Incubate for 5 min at room temperature (15–25°C). Centrifuge for 5 min at 3000–5000 x *g*.

11. Repeat step 10.