

| Product Information             |                |  |  |  |
|---------------------------------|----------------|--|--|--|
| ZipScript™ One-Step RT-qPCR Mix |                |  |  |  |
| Part Number                     | P7640L         |  |  |  |
| Storage Temperature             | -25°C to -15°C |  |  |  |
| Reaction Size                   | 1000 Reactions |  |  |  |
| Volume                          | 0.8 mL         |  |  |  |
| Lot Number                      |                |  |  |  |
| Reference Number                |                |  |  |  |

# **Product Description:**

The ZipScript One-Step RT-qPCR Mix is a highly sensitive and reproducible RT-qPCR solution optimized for real-time PCR. The 25X enzyme mix is accompanied by a 2X reaction buffer.

| Product Specifications |   |  |  |  |
|------------------------|---|--|--|--|
| Assay                  | RT-qPCR   |  |  |  |
| Specification          | Amplification of Test Lot within 1Ct of Reference Lot in a one-step RT-qPCR Assay |  |  |  |

# **Quality Control Analysis:**

The functionality of the ZipScript is evaluated by amplification of three mRNA transcripts in a one-step RT-qPCR assay. The amplification threshold (Ct) of the test lot is compared to a reference lot.

### Notes:

Enzyme components were tested prior to formulation of the master mix and found free of contaminating endonucleases and exonucleases. Enzyme purity was >99% as determined by SDS-PAGE and negligible *E.coli* genomic DNA contamination was confirmed by qPCR. Specific activity was verified for each enzyme pre-formulation.

## Supplied with:

ZipScript Reaction Buffer 1 (5X) B7641

#### **Limitations of Use**

This product was developed, manufactured, and sold for *in vitro* use only. The product is not suitable for administration to humans or animals. MSDS sheets relevant to this product are available upon request.



## **Protocol**

# **ZipScript One-Step RT-qPCR Reaction Setup:**

- 1. All components and reaction set up should be kept on ice.
- 2. Thaw the 2X ZipScript Reaction Buffer I completely and vortex for 3-5 seconds to mix thoroughly. Quick spin to collect contents if necessary.
- 3. Prepare primer/probe mix. A final concentration of 0.4- $0.9~\mu M$  for each primer and 0.1- $0.5~\mu M$  for probe is recommended. However, the optimal concentration for primers/probe needs to be empirically determined for each assay.
- 4. Determine the number of reactions to prepare, including No Template Controls (NTCs). Add 10% extra volume to compensate for the pipetting loss.
- 5. Follow the table below to set up the reaction mix. It is recommended to make a master mix to minimize variations and potential errors.

| Components                     | Volume/Rxn                              | Final Concentration |
|--------------------------------|---|---------------------|
| 2X ZipScript Reaction Buffer 1 | 10 μL                                   | 1X                  |
| Primer/Probe Mix               | XμL                                     | Variable            |
| 25X ZipScript Enzyme Mix       | 0.8 μL                                  | 1X                  |
| RNA Template                   | Variable                                | -                   |
| 50X ROX (optional)             | 0.4 μL                                  | 1X                  |
| Nuclease-Free Water            | To a final <b>20 μL</b> reaction volume | -                   |

6. Seal PCR plate and spin briefly to bring down reagents.

### **Thermal Cycling Conditions:**

Program the cycling conditions based on the recommendations below.

| Standard Cycling Program* Steps     | Temperature | Time      | Cycles |
|-------------------------------------|-------------|-----------|--------|
| Reverse Transcription               | 50°C        | 15 min    | 1      |
| Taq Activation/Initial Denaturation | 95°C        | 2 min     | 1      |
| Denaturation                        | 95°C        | 15 sec    | 40     |
| Annealing/Extension*                | 60°C        | 30-60 sec | .0     |

<sup>\*</sup> Cycling parameters can be modified (especially the annealing/extension condition) to fit specific primer/probe selection.