Quick-Start Protocol November 2016

DNeasy® PowerClean® Cleanup Kit

The DNeasy PowerClean Cleanup Kit can be stored at room temperature (15–25°C) until the expiry date printed on the box label.

Further information

- Safety Data Sheets: **www.qiagen.com/safety**
- Technical assistance: **support.qiagen.com**

Notes before starting

- Shake to mix Solution SB.
- If Solution SL has precipitated, heat at 60°C, gently inverting the tube periodically, until the precipitate has dissolved. Solution SL may be used while still warm.
- Wear gloves at all times.

1. Add up to 150 µl of DNA sample to a clean 2 ml collection tube (provided). If there is less than 150 µl of sample, adjust volume with distilled or deionized water.
2. Add 70 µl of Solution CU to DNA. Gently invert 3–5 times to mix.
3. Add 20 µl of Solution SL and invert 3–5 times to mix.
4. Add 85 µl of Solution AA and invert 3–5 times to mix. Incubate at 2–8°C for 5 min.
5. Centrifuge tubes at 10,000 x *g* for 1 min at room temperature.
6. Taking care to avoid the pellet, transfer the entire volume of the supernatant to a clean 2 ml collection tube (provided).
7. Add 70 µl of Solution IRS and invert 3–5 times to mix. Incubate at 2–8°C for 5 min.
8. Centrifuge tubes at 10,000 x *g* for 1 min at room temperature.
9. Taking care to avoid the pellet, transfer the entire volume of the supernatant to a clean 2 ml Collection Tube (provided).

10. Add 800 µl of Solution SB and vortex for 5 s.
11. Load approximately 600 µl onto an MB Spin Column and centrifuge at 10,000 x *g* for 1 min at room temperature. Discard flow through.
12. Add the remaining 600 µl supernatant to the MB Spin Column and centrifuge at 10,000 x *g* for 1 min at room temperature.
    Note: A total of two loads may be required for each sample processed.
13. Add 500 µl of Solution CB to the MB Spin Column and centrifuge at 10,000 x *g* for 30 s at room temperature. Discard flow through.
    Note: An additional wash of the MB Spin Column with 100% ethanol may be performed prior to the final dry spin. This may enhance the purity of the DNA obtained from some sample types. After discarding the flow through in Step 13, add 650 µl of 100% ethanol to the spin column and centrifuge at 10,000 x *g* for 30 s. To completely remove residual ethanol, discard the flow through and increase the dry spin (step 14) to 2 min at 13,000 x *g* or max speed. Continue with step 15.
14. Centrifuge the MB Spin Column at 13,000 x *g* for 1 min at room temperature.
15. Carefully place the MB Spin Column in new 2 ml collection tube (provided). Avoid splashing any Solution CB onto the MB Spin Column.
16. Add 50–100 µl of Solution EB (depending on starting volume of DNA sample) to the center of the white filter membrane. Incubate for 1 min at room temperature.
    Note: For efficient elution, use a **minimum** of 50 µl of Solution EB, irrespective of starting sample volume. By reducing elution volume, it is possible to obtain more concentrated DNA.
17. Centrifuge at 10,000 x *g* for 30 s at room temperature.
18. Discard the MB Spin Column. The DNA is now ready for downstream applications.
    Note: We recommend storing DNA frozen (–20° to –80°C) as Solution EB does not contain EDTA.