Quick-Start Protocol May 2019

# QIAprep® 96 *Plus* Miniprep Kit

The QIAprep 96 *Plus* Miniprep Kit (cat. no. 27291) and the QIAprep 96 *Plus* BioRobot® Kit (cat. no. 962241) can be stored at room temperature (15–25°C) for up to 9 months if not otherwise stated on label.

## Further information

- *QIAprep 96* Plus *Miniprep Handbook*: **www.qiagen.com/HB-1186**
- Safety Data Sheets: **www.qiagen.com/safety**
- Technical assistance: **support.qiagen.com**

## Notes before starting

- Add RNase A solution to Buffer P1, mix, and store at 2–8°C.
- Add ethanol (96–100%) to Buffer PE concentrate (see bottle label for volume).
- Assemble the QIAvac 96: Place an S-Block (square-well block) inside the QIAvac base. Assemble the QIAvac top plate with the TurboFilter® 96 plate and place over the base. The vacuum should be regulated to –300 mbar.

1. Completely resuspend pelleted bacteria from ≤5 ml LB culture in 300 µl Buffer P1.
2. Add 300 µl Buffer P2 to each well. Dry the top of the S-Block thoroughly with a paper towel, and seal the block with tape. Gently invert the block 6–8 times or until the solution becomes viscous and clear. Incubate at room temperature (15–25°C) for up to 5 min.
3. Remove tape from block. Add 300 µl Buffer S3 to each well. Thoroughly dry top of S-Block with paper towel, and seal block with new tape. Mix by gently inverting block 6–8 times.
4. Remove the tape from the block. Transfer the lysates from step 3 into the wells of TurboFilter 96 plate on the assembled QIAvac 96. Apply vacuum until all samples have passed through the wells of the TurboFilter 96 plate into a fresh S-Block.
5. After all liquid has been drawn through the TurboFilter 96 plate, switch off vacuum and ventilate the QIAvac 96 slowly.

6. Add 300 µl Buffer BB to the cleared lysate in each well of S-Block. Dry the top of the S-Block thoroughly with paper towel, seal with new tape, and invert 1–2 times to mix.
7. Place a waste tray into the base of the QIAvac 96. Place the QIAvac 96 top plate over the base. Place the Plasmid *Plus* 96 plate into top plate. Transfer lysates from the S-Block to the Plasmid *Plus* 96 plate. Apply vacuum until all samples have passed through.
8. To wash the DNA, add 900 µl Buffer PE to each well of the Plasmid *Plus* 96 plate. Apply vacuum until all samples have passed through.
9. Empty the waste tray of the QIAvac 96, put it back in, and apply maximum vacuum for 10 min. Switch off vacuum and ventilate the QIAvac 96 slowly. Lift the top plate from the base (but not the Plasmid *Plus* 96 plate from the top plate), vigorously tap the top plate on a stack of absorbent paper until no more drops come out, and blot the nozzles of the Plasmid *Plus* 96 plate with clean absorbent paper.
10. To elute the DNA from the plate, replace the waste tray with the Elution Microtube Adapter or an empty 96-well microplate.
11. Place the Elution Microtube Rack containing Elution Microtubes onto the adapter.
12. Place top plate back on base, making sure the Plasmid *Plus* 96 plate is seated securely.
13. Add 80 µl Buffer EB to the center of each well of the Plasmid *Plus* 96 plate.
14. Let stand for 3 min, and then apply maximum vacuum for 1 min.
15. Switch off vacuum and ventilate the QIAvac 96 slowly.

## Revision History

| Document | Changes | Date |
|---|---|---|
| HB-0598-003 | Replaced flat-bottom blocks with S-Blocks. | May 2019 |

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