Quick-Start Protocol April 2016

# DNeasy *mericon*® 96 QIAcube HT Kit, Part 2

Store DNeasy 96 plates and all buffers at room temperature (15–25°C). QIAGEN® Proteinase K is stable for at least 1 year after delivery when stored at room temperature if not otherwise stated on label. For storage for >1 year, or if ambient temperatures often exceed 25°C, store QIAGEN Proteinase K solution at 2–8°C.

## Further information

- *DNeasy* mericon *96 QIAcube HT Handbook*: **www.qiagen.com/HB-1592**
- Safety Data Sheets: **www.qiagen.com/safety**
- Technical assistance: **support.qiagen.com**

## Notes before starting

- Refer to part 1of this protocol before proceeding further.

## DNA purification – food and plant samples

12. Place 300 mg homogenized food or plant sample in a 2 ml microcentrifuge tube and add 1.5 ml Food Lysis Buffer and 4 µl proteinase K solution. Vortex briefly to ensure complete mixing and distribution of the sample material.
13. Incubate for 30 min at 60°C with constant shaking. To enhance inhibitor precipitation, cool the sample to room temperature (15–25°C) on ice after incubation.
14. Centrifuge for 5 min at 2500 x *g*. **IMPORTANT**: Keep the supernatant.
15. Pipet 400 µl chloroform into a fresh 2 ml microcentrifuge tube.

16. Carefully transfer 550 µl of the clear supernatant from step 14 to the microcentrifuge tube containing the chloroform. Be sure not to carry over material from the bottom phase, which contains precipitated food debris. Vortex the microcentrifuge tube from step 15 vigorously for 15 s and centrifuge at 14,000 x *g* for 15 min.
17. Transfer the indicated volumes of all reagents into the corresponding reagent troughs, close the lids and place them on the indicated positions on the worktable.
18. After sample centrifugation, transfer 350 µl of the upper, aqueous phase to the selected S-Block wells. Place the S-Block in the B1 position of the worktable.

## Automated sample processing

19. Start the run immediately and perform the pre-run check.
20. After completing the pre-run check, close the instrument hood and click **OK**. Click **Cancel** when the **Save as** dialog box appears. The protocol run begins.
21. Cover the elution plate (EMTR) with the lid and remove from the elution chamber, when the protocol is complete.

    **Note**: Two liquid phases might be found in the Elution Microtubes. If this is the case, TopElute Fluid will be found as a top layer over the elution buffer. It is inert and has no effect on downstream applications.
22. Discard used plasticware. We recommend discarding leftover reagents in the reagent troughs.
23. Clean the carriage, channeling block, channeling block holder and tip chute. Turn on the UV lamp to decontaminate the worktable.

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