

February 2018

QIASymphony[®] RGQ Application Sheet

artus[®] HSV-1/2 QS-RGQ Kit (sample type: CSF)

R2

IVD

CE

REF

4500363 *artus* HSV-1/2 QS-RGQ Kit, version 1



Check availability of new electronic labeling revisions at www.qiagen.com/products/artushsv-12pcrkitce.aspx before test execution.

General Information

Kit	<i>artus</i> HSV-1/2 QS-RGQ Kit, Version 1 (cat.no. 4500363)
Validated sample material	CSF
Front-end purification	QIAasymphony DSP Virus/Pathogen Mini Kit (cat. no. 937036)
Sample volume (including excess volume)	350 µl
Assay Parameter Set	<i>artus</i> _HSV_CSF200_V5
Default Assay Control Set	Cellfree200_V7_DSP_ <i>artus</i> _HSV
Elution volume	60 µl
Required software version	Version 4.0 or higher
Master mix volume	30 µl
Template volume	20 µl
Number of reactions	4–24
Runtime on AS module	For 4 reactions: approximately 9 minutes For 72 reactions: approximately 35 minutes

Materials Required but Not Provided

Purification kit

- QIASymphony DSP Virus/Pathogen Mini Kit (cat. no. 937036)

Adapters for the QIASymphony SP

- Elution Microtube Rack QS (Cooling Adapter, EMT, v2, Qsym, cat. no. 9020730)
- Transfer frame
- Tube Insert 3B (Insert, 2.0ml v2, samplecarr. (24), Qsym, cat. no. 9242083)

Consumables for the QIASymphony SP

- Sample Prep Cartridges, 8-well (cat. no. 997002)
- 8-Rod Covers (cat. no. 997004)
- Filter-Tips, 1500 µl (cat. no. 997024)
- Filter-Tips, 200 µl (cat. no. 990332)
- Elution Microtubes CL (cat. no. 19588)
- Tip disposal bags (cat. no. 9013395)
- Micro tubes 2.0 ml Type H or Micro tubes 2.0 ml Type I (Sarstedt®, cat. nos. 72.693 and 72.694, www.sarstedt.com) for use with samples and internal controls

Adapters and reagent holders for the QIASymphony AS

- Reagent holder 1 QS (Cooling Adapter, Reagent Holder 1, Qsym, cat. no. 9018090)
- RG Strip Tubes 72 QS (Cooling Adapter, RG Strip Tubes 72, Qsym, cat. no. 9018092)

Consumables for the QIASymphony AS

- Strip Tubes and Caps, 0.1 ml (cat. no. 981103)
- Tubes, conical, 2 ml, Qsym AS (cat. no. 997102) or Micro tubes 2.0 ml Type I (Sarstedt, cat. no. 72.694.005)
- Alternatively: Tubes, conical, 5 ml, Qsym AS (cat. no. 997104) or Tubes with flat base from PP (Sarstedt, cat. no. 60.558.001)
- Filter-Tips, 1500 µl (cat. no. 997024)
- Filter-Tips, 200 µl (cat. no. 990332)
- Filter-Tips, 50 µl (cat. no. 997120)
- Tip disposal bags (cat. no. 9013395)

Specimen Handling and Storage

Sample collection	CSF
Sample storage	Samples should be stored at 2–8°C, or at –20°C
Sample transport	Shatterproof transport Shipment within 24 hours Mail shipment according to legal instructions for the transport of pathogen material*
Sample preparation	Prevent formation of foam in or on the samples Samples should be equilibrated to room temperature (15–25°C) before starting the run.

* International Air Transport Association (IATA). Dangerous Goods Regulations.

Procedure

Preparation of carrier RNA and addition of the internal control to the samples

Using the QIAasymphony DSP Virus/Pathogen Mini Kit in combination with the *artus* HSV-1/2 QS-RGQ Kit requires introduction of the internal control (HSV-1/2 RG IC) into the purification procedure to monitor the efficiency of sample preparation and downstream assay.

Internal controls must be added with carrier RNA (CARRIER)–Buffer AVE (AVE) mixture, and the total volume of the internal control–carrier RNA (CARRIER)–Buffer AVE (AVE) mixture remains 120 µl.

The table represents the addition of internal control to the isolation at a ratio of 0.1 µl per 1 µl elution volume. We recommend preparing fresh mixtures for each run just before use.

Alternatively, the “IC Calculator” Tool in the QIAasymphony Management Console may be used.

Component	Volume (µl) (Sarstedt tubes)*	Volume (µl) (Corning tubes)†
Stock carrier RNA (CARRIER)	2.5	2.5
Internal control‡	9	9
Buffer AVE	108.5	108.5
Final volume per sample (excluding dead volume)	120	120
Total volume for n samples	$(n \times 120) + 360^{\S}$	$(n \times 120) + 600^{\P}$

* Micro tubes 2.0 ml Type H and Micro tubes 2.0 ml Type I, Sarstedt cat. nos. 72.693 and 72.694.

† Tubes 14 ml, 17 x 100 mm polystyrene round-bottom (Corning® Inc., cat. no. 352051; Becton Dickinson was the previous supplier of this tube and Corning Inc. is now the new supplier).

‡ The calculation of the amount of internal control is based on the initial elution volumes (90 µl). Additional void volume depends on the type of sample tube used.

§ Internal control mixture corresponding to 3 additional samples (i.e., 360 µl) is required. Do not fill more than 1.92 ml total volume (corresponding to a maximum of 13 samples. These volumes are specific for Micro tubes 2.0 ml Type H and Micro tubes 2.0 ml Type I, Sarstedt cat. nos. 72.693 and 72.694).

¶ Internal control mixture corresponding to 5 additional samples (i.e., 600 µl) is required. Do not fill more than 13.92 ml total volume (corresponding to a maximum of 111 samples. These volumes are specific for Tubes 14 ml, 17 x 100 mm polystyrene round-bottom, Corning Inc., cat. no. 352051; Becton Dickinson was the previous supplier of this tube and Corning Inc. is now the new supplier).

QIAsymphony SP setup

“Waste” drawer

Unit box holder 1–4	Empty unit boxes
Waste bag holder	Waste bag
Liquid waste bottle holder	Empty and install liquid waste bottle

“Eluate” drawer

Elution rack	Elution Microtubes CL in Elution Microtube Rack QS and Transfer frame Use slot 1, cooling position
Elution volume*	Preselected elution volume: 60 µl Initial elution volume: 90 µl

* The elution volume is preselected for the protocol. This is the minimum accessible volume of eluate in the final elution tube. The initial volume of elution solution is required to ensure that the actual volume of eluate is the same as the preselected volume.

“Reagents and Consumables” drawer

RC Position 1 and 2	Load 1 reagent cartridge (RC) for up to 96 samples
Tip rack holder positions 1–18	Load sufficient racks of disposable filter-tips, 200 µl and 1500 µl (see “Required plasticware for 1–4 sample batches”, page 7)
Unit box holder positions 1–4	Load unit boxes containing sample prep cartridges and 8-Rod Covers (see “Required plasticware for 1–4 sample batches”, page 7)

“Sample” drawer

Sample type	CSF
Sample volume (including excess volume)	350 µl
Sample tubes	Micro tubes 2.0 ml Type H or Micro tubes 2.0 ml Type I (Sarstedt, cat. nos. 72.693 and 72.694)
Insert	Tube Insert 3B (cat. no. 9242083)

Required plasticware for 1–4 sample batches

Component	One batch, 24 samples*	Two batches, 48 samples*	Three batches, 72 samples*	Four batches, 96 samples*
Disposable filter-tips, 200 µl†‡	30	54	78	102
Disposable filter-tips, 1500 µl†‡	101	182	271	354
Sample prep cartridges§	21	42	63	84
8-Rod Covers¶	3	6	9	12

* Use of more than one internal control tube per batch and performing more than one inventory scan requires additional disposable filter tips.

† There are 32 filter-tips/tip rack.

‡ Number of required filter-tips includes filter-tips for 1 inventory scan per reagent cartridge.

§ There are 28 sample prep cartridges/unit box.

¶ There are twelve 8-Rod Covers/unit box.

QIAsymphony AS setup

Consumables

During the setup, the appropriate positions for each consumable on the QIAsymphony AS module are indicated on the touchscreen of the instrument.

Consumable	Name on touchscreen	For use with adapter/ reagent holder
Strip Tubes and Caps, 0.1 ml (250)	QIA#981103 *StripTubes 0.1	RG Strip Tubes 72 QS
Tubes, conical, 2 ml, Qsym AS (500) ^{†‡}	QIA#997102 *T2.0 ScrewSkirt [§]	Reagent holder 1 QS
Tubes, conical, 5 ml, Qsym AS (500) ^{†‡}	QIA#997104 *T5.0 ScrewSkirt [§]	Reagent holder 1 QS

* Indicates labware that can be cooled using a cooling adapter with bar code.

[†] For master mix components, system-prepared master mix, assay standards, and assay controls.

[‡] Alternatively, the Sarstedt tubes described in "Material Required but Not Provided", page 3, can be used.

[§] The suffix "(m)" in the touchscreen indicates that liquid level calculations for the respective tube have been optimized for reagents forming a concave meniscus.

Adapters and reagent holders

Rack/reagent holder	Name	Number required [¶]
Reagent holders	Reagent holder 1 QS	1
Sample racks	RG Strip Tubes 72 QS	1

[¶] Calculated for an assay run with 72 reactions.

Filter-tips

Load tip racks starting with tip slots 1, 2 and 3 in the "Eluate and Reagents" drawer, and then load tip racks into tip slots 7, 8 and 9 in the "Assays" drawer.

Consumable	Name on touchscreen	Minimum number for 24 reactions	Minimum number for 72 reactions
Filter-Tips, 1500 µl (1024)	1500 µl	4	6
Filter-Tips, 200 µl (1024)	200 µl	9	8
Filter-Tips, 50 µl (1024)	50 µl	25	73
Tip Disposal Bags	–	1	1

PCR on the Rotor-Gene Q*

Please refer to the software-specific protocol sheet *Settings to run artus QS-RGQ Kits* at www.qiagen.com/products/artushsv-12pcrkite.aspx for protocol details.

Specific settings for the *artus HSV-1/2 QS-RGQ Kit*

With Rotor-Gene® software 2.1 or higher, the specific settings are shown below.

Reaction Volume (µl)	50
Hold	Hold Temperature: 95 degrees Hold Time: 10 minutes
Cycling	45 times 95 degrees for 15 seconds 65 degrees for 30 seconds (Acquire on Green, Orange, Yellow and activate touchdown function for 10 cycles) 72 degrees for 20 seconds
Auto-Gain Optimisation Setup	65 degrees (Samples: Green, Orange; IC: Yellow)

Interpretation of results

This section describes interpretation of results on the Rotor-Gene Q. Review also the sample status information from the QIAasymphony SP/AS result files for analysis of the complete sample-to-result workflow. Only samples with a valid status should be used.

The *artus HSV-1/2 QS-RGQ Kit* can be run on the Rotor-Gene Q using manual analysis with Rotor-Gene Q software 2.1 or higher. The following sections describe interpretation of results using Rotor-Gene Q software 2.1 or higher.

* If applicable, Rotor-Gene Q 5plex HRM instrument with a production date of January 2010 or later. The production date can be obtained from the serial number on the back of the instrument. The serial number is in the format "mmyynn" where "mm" indicates the production month in digits, "yy" indicates the last two digits of the production year, and "nnn" indicates the unique instrument identifier.

Signal detection and conclusions

Signal in channel Cycling Green	Signal in channel Cycling Orange	Signal in channel Cycling Yellow	Result	Interpretation
Yes	No	Yes*	Positive	The sample contains HSV-1 DNA. No HSV-2 DNA is detectable. [†]
No	Yes	Yes*	Positive	The sample contains HSV-2 DNA. No HSV-1 DNA is detectable. [†]
Yes	Yes	Yes*	Positive	The sample contains HSV-1 and HSV-2 DNA.
No	No	Yes	Negative	No HSV-1/2 DNA is detectable. [†]
No	No	No	Inconclusive	No result can be concluded. [‡]

* Signal is dispensable, since high initial concentrations of HSV-1 and/or HSV-2 DNA can lead to reduced or absent fluorescence signal in the Cycling Yellow channel (competition).

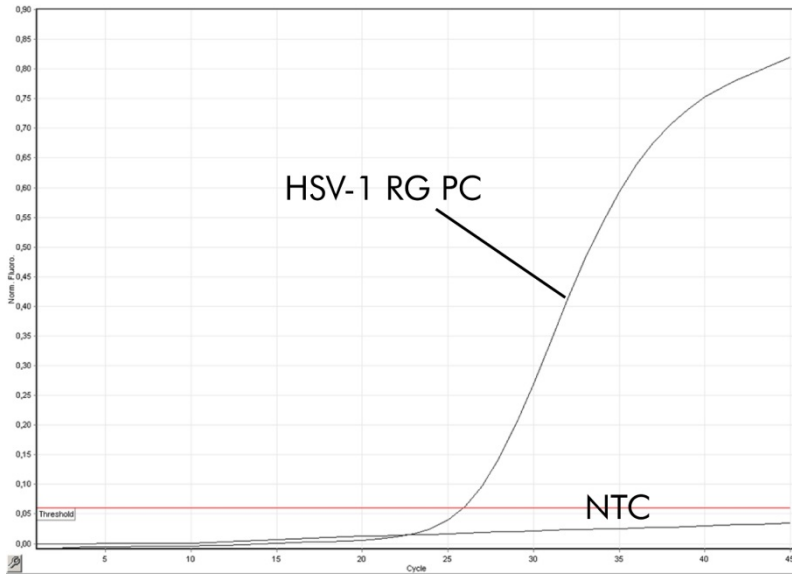
[†] If the C_T value for the internal control of a negative sample is more than 3 cycles higher than the C_T value for the internal control of the no template control in the run ($C_{T\text{ IC Sample}} - C_{T\text{ IC NTC}} > 3$), then the sample should be treated as invalid. No result can be concluded.

[‡] Information regarding error sources and their solution can be found in "Troubleshooting Guide" of the *artus HSV-1/2 QS-RGQ Kit Handbook*.

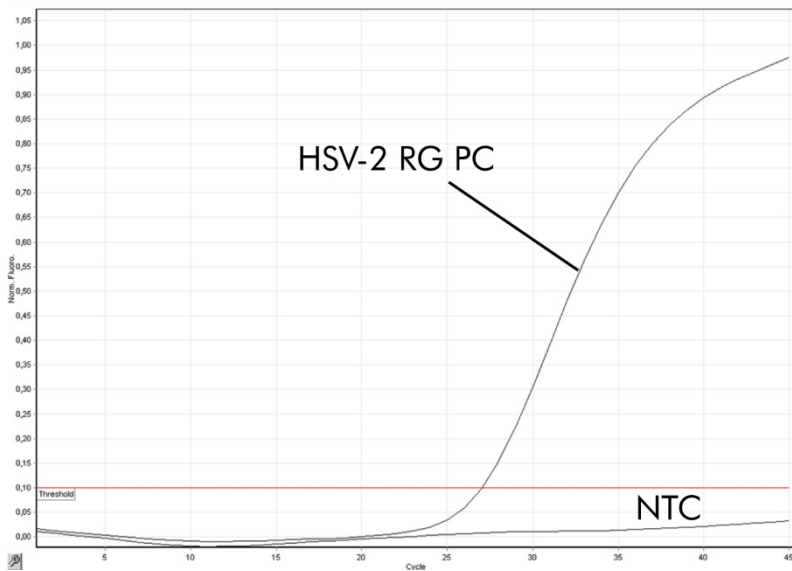
Threshold setup for the PCR analysis

The optimal threshold settings for a given combination of Rotor-Gene Q instrument and *artus* QS-RGQ Kit should be set empirically by testing each individual combination since it is a relative value depending on the overall diagnostic workflow. The threshold can be set at a preliminary value of 0.04 for the analysis of the first PCR run, but this value should be fine-tuned in a comparative analysis of the next runs of the workflow. The threshold should be set manually just above the background signal of the negative controls and negative samples. The mean threshold value calculated from these experiments will most likely work for the majority of future runs, but the user should nevertheless review the generated threshold value at regular intervals. The threshold value will usually be in the range of 0.03–0.05 and should be rounded to no more than three decimal places.

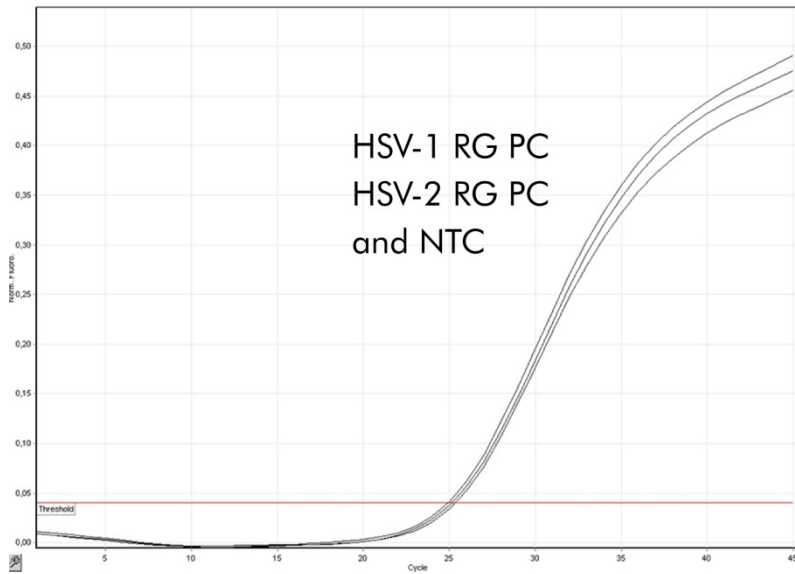
Examples of positive and negative PCR reactions



Detection of the HSV-1 positive control (HSV-1 RG PC) in fluorescence channel Cycling Green. NTC: No template control (negative control).



Detection of the HSV-2 positive control (HSV-2 RG PC) in fluorescence channel Cycling Orange. NTC: No template control (negative control).



Detection of the internal control (IC) in fluorescence channel Cycling Yellow with simultaneous amplification of the positive controls (HSV-1 RG PC and HSV-2 RG PC). NTC: No template control.

Document revision history

R2, February 2018	Removed footnote regarding setup of 216 assays. Changed to new versions of QIASymphony protocols. Updated required materials for setup of maximum 72 reactions. Added information on use of QMC "IC Calculator" tool. Updated naming of Corning labware (previously Becton Dickinson). Added specific run settings for Rotor-Gene Q (use of touchdown function, acquisitions). Removed reference to Rotor-Gene AssayManager®.
-------------------	---

For up-to-date licensing information and product-specific disclaimers, see the respective QIAGEN kit handbook or user manual. QIAGEN kit handbooks and user manuals are available at www.qiagen.com or can be requested from QIAGEN Technical Services or your local distributor.

Trademarks: QIAGEN®, Sample to Insight®, QIASymphony®, *artus*®, Rotor-Gene®, Rotor-Gene AssayManager® (QIAGEN Group); Corning® (Corning Inc.); Sarstedt® (Sarstedt AG and Co.). Registered names, trademarks, etc. used in this document, even when not specifically marked as such, are not to be considered unprotected by law.
02/2018 HB-0403-S01-002 © 2012–2018 QIAGEN, all rights reserved.

