

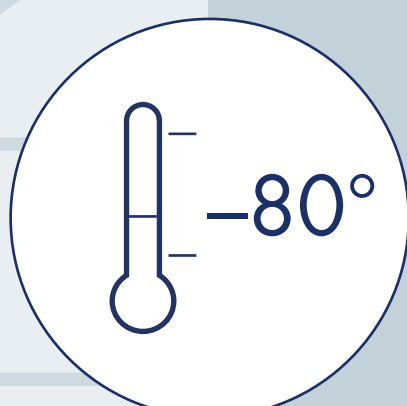
The DO's and DON'Ts of working with RNA

Handling, storage and stabilization

DO

preserve the expression profile of RNA by:

- keeping RNA samples frozen in liquid nitrogen or at -80°C
- alternatively, use stabilization reagents like RNAlater®



DON'T

- let frozen samples thaw

DO

- designate a special workbench for RNA
- clean the bench with cleaning solutions like RNaseZap®



DON'T

- let dust particles settle on the benchtop

DO

- wear disposable gloves
- Did you know that RNases are secreted from your skin?
- wear a mask while working



DON'T

- work with bare hands
- breathe on your samples

DO

- use sterile, disposable plasticware and DEPC-treated reagents and glassware
- treat non-disposable plasticware with 0.1 M NaOH/1 mM EDTA and RNase-free water
- clean electrophoresis tanks with detergent (0.5% SDS) and pipettors, gel boxes and combs with RNaseZap



DON'T

- reuse autoclaved pipette tips

DO

- perform dry, cryogenic tissue disruption
- perform homogenization in lysis buffer under precooled conditions
- remove proteins bound to RNA by treating sample with a protease or by vortexing
- rinse your homogenizer with lysis buffer between samples to minimize cross-contamination

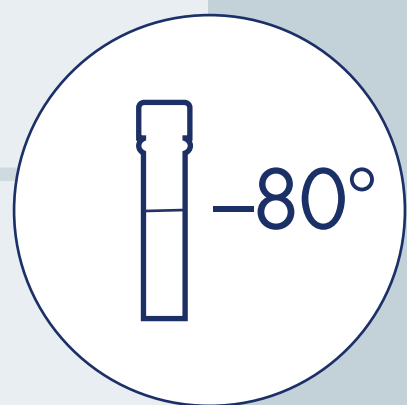


DON'T

- rinse RNAlater from tissues before processing

DO

- store purified RNA, aliquoted in RNase-free water at -20°C or -80°C (under these conditions, no degradation of RNA is detectable after 1 year)



DON'T

- resuspend RNA in DEPC water; residual DEPC can interfere with downstream reactions

DO

- follow protocols with short incubation times on ice and keep tubes closed whenever possible



DON'T

- overdry the RNA pellet
- deviate from suggested time intervals

Find your RNA stabilization and purification solution for any sample type at www.qiagen.com/RNA.

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