

# QIASymphony SP Protocol Sheet

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## Purification of RNA from FFPE Sections Using RNA\_FFPE\_130\_V7

### General information

This protocol is for purification of total RNA from FFPE tissue sections using the QIASymphony® SP and the QIASymphony RNA Kit. The protocol recovers total RNA, including small RNA fragments. However, the protocol is not fully optimized for the isolation of miRNA. Depending on the type of tissue, miRNA recoveries may be lower than expected.

For purification of total RNA from biopsy needle core punches, go to the “Resources” tab at [www.qiagen.com/RNeasyFFPEKit](http://www.qiagen.com/RNeasyFFPEKit) and select the relevant protocol from the list.

**Note:** It is the user’s responsibility to validate performance using this combination for any procedures used in their laboratory.

<b>Kit</b>	QIASymphony RNA Kit (cat. no. 931636)
<b>Sample material</b>	FFPE tissue samples 5–20 $\mu\text{m}$ thick
<b>Protocol name</b>	RNA_FFPE_130_V7
<b>Default Assay Control Set</b>	ACS_RNA_FFPE_130_V7
<b>Editable</b>	Elution volume: 50 $\mu\text{l}$ , 100 $\mu\text{l}$ , 200 $\mu\text{l}$
<b>Required software version</b>	Version 4.0

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## “Sample” drawer

<b>Sample type</b>	FFPE microtome sections
<b>Sample amount</b>	Lysate prepared from 1–2 sections 5–20 $\mu\text{m}$ thick
<b>Lysate volume</b>	130 $\mu\text{l}$
<b>Primary sample tubes</b>	n/a
<b>Secondary sample tubes</b>	We recommend using 2 ml tubes (e.g., Sarstedt® cat. no. 72.693 or 72.608) or S-Blocks (cat. no. 19585). For a full list of compatible vessels, see <a href="http://www.qiagen.com/QIASymphony/Resources">www.qiagen.com/QIASymphony/Resources</a>
<b>Inserts</b>	For more information, see the “Resources” tab at <a href="http://www.qiagen.com/QIASymphonyRNAKit">www.qiagen.com/QIASymphonyRNAKit</a>

n/a = not applicable.

## “Reagents and Consumables” drawer

<b>Position A1 and/or A2</b>	Reagent cartridge
<b>Position B1</b>	n/a
<b>Tip rack holder 1–17</b>	Disposable filter-tips, 200 $\mu\text{l}$ or 1500 $\mu\text{l}$
<b>Unit box holder 1–4</b>	Unit boxes containing sample prep cartridges or 8-Rod Covers
<b>Tip racks slots 5 and 12</b>	Accessory troughs for ethanol

n/a = not applicable.

## “Waste” drawer

<b>Unit box holder 1–4</b>	Empty unit boxes
<b>Waste bag holder</b>	Waste bag
<b>Liquid waste bottle holder</b>	Empty liquid waste bottle

## “Eluate” drawer

Elution rack (we recommend using slot 1, cooling position)

For more information, see the “Resources” tab at [www.qiagen.com/QIASymphonyRNAKit](http://www.qiagen.com/QIASymphonyRNAKit)

## Required plasticware

	24 samples	96 samples
Reagent cartridges	1	2 <sup>§</sup>
Sample prep cartridges*	15	45
8-Rod Covers <sup>†</sup>	3	9
Disposable filter-tips, 1500 $\mu$ l <sup>‡</sup>	92	276
Disposable filter-tips, 200 $\mu$ l <sup>‡</sup>	24	96
Ethanol (ml)	140	2 x 140

\* 28 sample prep cartridges/unit box.

<sup>†</sup> Twelve 8-Rod Covers/unit box.

<sup>‡</sup> 32 filter-tips/tip rack; the inventory scan requires additional tips (two 200  $\mu$ l and seven 1500  $\mu$ l tips).

<sup>§</sup> 72 samples per reagent cartridge.

**Note:** Numbers of filter-tips given may differ from the numbers displayed in the touchscreen depending on settings. We recommend loading the maximum possible number of tips.

## Elution volume

The elution volume is selected in the touchscreen. Depending on the sample type and RNA content, the final eluate volume may vary by up to 15  $\mu$ l less than the selected volume. Elution in smaller volumes increases the final RNA concentration, but reduces the yield and increases variability of the eluate volume. We recommend using the smallest elution volume only where the intended downstream application requires a higher RNA concentration.

## Preparation of sample material

When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, consult the appropriate material safety data sheets (MSDSs), available from the product supplier.

### Important points before starting

- Deparaffinization Solution (cat. no. 19093), QIAGEN Proteinase K (cat. no. 19131), Buffer PKD (cat. no. 1034963), and DNase Booster Buffer (cat. no. 1064143) are required for the RNA\_FFPE\_130\_V7 protocol, but are not supplied with the QIASymphony RNA Kit. They should be ordered separately.

### Things to do before starting

- Transfer 1.4 ml of DNase solution to each of the tubes in positions 1 and 2 of the enzyme rack on the reagent cartridge. For more information about preparation of DNase I, see the *QIASymphony RNA Handbook*, page 25.
- In the RNA\_FFPE\_130\_V7 protocol, proteinase K is added in the manual pretreatment of the samples. Therefore, tubes in positions 3 and 4 can remain empty with the lids on.
- Transfer 2 ml DNase Booster Buffer to the tube in position 5 of the enzyme rack on the reagent cartridge.
- Set a thermal mixer or heated orbital incubator to 56°C for use in step 6.

### Pretreatment protocol for FFPE sections

1. **Using a scalpel, trim excess paraffin off the sample block.**
2. **Cut sections 5–20 µm thick.**
3. **If the sample surface has been exposed to air, discard the first 2–3 sections.**
4. **Immediately place the sections in a 2 ml sample tube compatible with the sample carrier of the QIASymphony SP (not supplied).**
5. **Add 160 µl Deparaffinization Solution, vortex vigorously for 10 s, and centrifuge briefly to bring the sample to the bottom of the tube.**

Deparaffinization Solution is not supplied with the QIASymphony RNA Kit and should be ordered separately (cat. no. 19093).

6. **Incubate at 56°C for 3 min, then allow to cool at room temperature.**

If samples turn waxy or solid upon cooling, add additional Deparaffinization Solution and repeat the 56°C incubation.

**7. Add 120  $\mu$ l Buffer PKD, and mix by vortexing.**

Buffer PKD is not supplied with the QIASymphony RNA Kit and should be ordered separately (cat. no. 1034963).

**8. Centrifuge for 1 min at 11,000 x g (10,000 rpm).**

**9. Add 10  $\mu$ l proteinase K to the lower, uncolored phase. Mix gently by pipetting up and down.**

**10. Incubate at 56°C for 15 min.**

**Note:** Some particulate non-dissolved matter may remain after this step. It is not required to completely digest all sample material in order to achieve maximum RNA yields.

**11. Place the tubes containing the digested samples into the appropriate sample carrier, and load them into the "Sample" drawer.**

**12. Begin the purification process, as described in the "General Purification Protocol". See the QIASymphony RNA Handbook, page 24.**

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