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RNeasy[®] MinElute[®] Cleanup Handbook

For RNA cleanup and concentration with small elution volumes

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Kit Contents

RNeasy MinElute Cleanup Kit	(50)
Catalog no.	74204
Number of preps	50
RNeasy MinElute Spin Columns (each in a 2 ml Collection Tube)	50
Collection Tubes (1.5 ml)	50
Collection Tubes (2 ml)	100
Buffer RLT*	45 ml
Buffer RPE† (concentrate)	11 ml
RNase-Free Water	10 ml
Quick-Start Protocol	1

* Contains a guanidine salt. Not compatible with disinfecting reagents containing bleach. See page 4 for safety information.

† Before using for the first time, add 4 volumes of ethanol (96–100%) as indicated on the bottle to obtain a working solution.

Storage

Store RNeasy MinElute spin columns immediately upon receipt at 2–8°C. Store the remaining components of the RNeasy MinElute Cleanup Kit dry at room temperature (15–25°C). All components of the kit are stable for at least 9 months under these conditions, if not otherwise stated on the label.

Intended Use


The RNeasy MinElute Cleanup Kit is intended for molecular biology applications. This product is not intended for the diagnosis, prevention or treatment of a disease. All due care and attention should be exercised in the handling of the products.

QIAcube® Connect is designed to perform fully automated purification of nucleic acids and proteins in molecular biology applications. The system is intended for use by professional users trained in molecular biological techniques and the operation of QIAcube Connect.

We recommend all users of QIAGEN® products to adhere to the NIH guidelines that have been developed for recombinant DNA experiments or to other applicable guidelines.

Safety Information

When working with chemicals, always wear a suitable lab coat, disposable gloves and protective goggles. For more information, please consult the appropriate safety data sheets (SDSs). These are available online in convenient and compact PDF format at www.qiagen.com/safety where you can find, view and print the SDS for each QIAGEN kit and kit component.

<p>CAUTION</p> 	<p>CAUTION: DO NOT add bleach or acidic solutions directly to the sample-preparation waste.</p>
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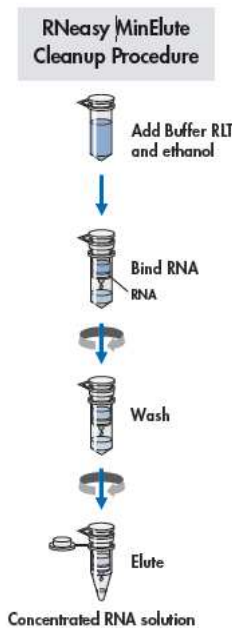
Buffer RLT contains guanidine thiocyanate, which can form highly reactive compounds when combined with bleach. If liquid containing this buffer is spilt, clean with suitable laboratory detergent and water. If the spilt liquid contains potentially infectious agents, clean the affected area first with laboratory detergent and water, and then with 1% (v/v) sodium hypochlorite.

Quality Control

In accordance with QIAGEN's ISO-certified Quality Management System, each lot of RNeasy MinElute Cleanup Kit is tested against predetermined specifications to ensure consistent product quality.

Introduction

The RNeasy MinElute Cleanup Kit uses novel technology to purify and concentrate RNA. RNA amounts from 45 µg down to a few picograms, which correspond to less than 1 cell, can be purified from enzymatic reactions (e.g., *in vitro* transcription, DNase digestion, RNA labeling) or from RNA samples requiring desalting (e.g., after phenol extraction and ethanol precipitation). The kit allows minimal elution volumes, making it well-suited for concentration of RNA (e.g., after RNA purification with the RNeasy Mini Kit or PAXgene™ Blood RNA Kit). QIAGEN provides a wide range of other kits for purification of total RNA from different sample sources; for details, visit www.qiagen.com/RNA.



Principle and procedure

RNeasy MinElute technology combines the selective binding properties of a silica-based membrane with the speed of microspin technology. Guanidine-thiocyanate-containing lysis buffer and ethanol are added to the sample to create conditions that promote selective binding of RNA to the silica membrane of the RNeasy MinElute spin column. The sample is then applied to the RNeasy MinElute spin column. RNA binds to the silica membrane, contaminants are efficiently washed away, and high-quality RNA is eluted in RNase-free water (see flowchart).

With the RNeasy MinElute procedure, all RNA molecules longer than 200 nucleotides are purified. The procedure provides an enrichment for mRNA since most RNAs <200 nucleotides (such as 5.8S rRNA, 5S rRNA and tRNAs, which together comprise 15–20% of total RNA) are selectively excluded. The size distribution of the purified RNA is comparable to that obtained by centrifugation through a CsCl cushion, where small RNAs do not sediment efficiently.*

Automated purification of RNA on QIAcube Instruments

Purification of RNA can be fully automated on QIAcube Connect or the classic QIAcube. The innovative QIAcube instruments use advanced technology to process QIAGEN spin columns, enabling seamless integration of automated, low-throughput sample prep into your laboratory workflow. Sample preparation using QIAcube instruments follows the same steps as the manual procedure (i.e., lyse, bind, wash and elute), enabling you to continue using the RNeasy MinElute Cleanup Kit for purification of high-quality RNA.

QIAcube instruments are preinstalled with protocols for purification of plasmid DNA, genomic DNA, RNA, viral nucleic acids and proteins, plus DNA and RNA cleanup. The range of

* For purification of miRNA and total RNA from a wide range of cells and tissues, we recommend using miRNeasy Kits. For details, visit www.qiagen.com/miRNA.

protocols available is continually expanding, and additional QIAGEN protocols can be downloaded free of charge at www.qiagen.com/qiacubeprotocols.



QIAcube Connect.

Equipment and Reagents to Be Supplied by User

When working with chemicals, always wear a suitable lab coat, disposable gloves and protective goggles. For more information, consult the appropriate safety data sheets (SDSs), available from the product supplier.

For all protocols

- Sterile, RNase-free pipet tips
- Microcentrifuge (with rotor for 2 ml tubes)
- Ethanol (80% and 96–100%)*
- Disposable gloves
- **Optional:** 14.3 M β -mercaptoethanol (β -ME) (commercially available solutions are usually 14.3 M) or, alternatively, 2 M dithiothreitol (DTT) in water

For concentration of RNA purified using the PAXgene Blood RNA Kit

- Elution Buffer BR5 (included in the PAXgene Blood RNA Kit)
- Heating block or water bath capable of reaching 65°C

* Do not use denatured alcohol, which contains other substances, such as methanol or methylethylketone.

Protocol: RNA Cleanup and Concentration

This protocol is designed for cleaning up RNA from enzymatic reactions, for desalting RNA samples and for concentrating RNA isolated by various methods. For concentration of total cellular RNA purified using the PAXgene Blood RNA Kit, follow the protocol on page 14.

Determining the correct amount of starting material

A maximum of 45 µg RNA in a maximum volume of 200 µl can be cleaned up in this protocol. This amount corresponds to the binding capacity of the RNeasy MinElute spin column. Do not overload the column as this will significantly reduce RNA yield and purity.

Important points before starting

- If preparing RNA for the first time, read Appendix A (page 21).
- Generally, DNase digestion is not required since RNeasy MinElute silica-membrane technology efficiently removes most of the DNA without DNase treatment. However, further DNA removal may be necessary for certain RNA applications that are sensitive to very small amounts of DNA (e.g., TaqMan® RT-PCR analysis with a low-abundance target). In these cases, DNA can be removed by a DNase digestion before starting RNA cleanup (see Appendix C, page 28).
- Buffer RLT contains a guanidine salt and is therefore not compatible with disinfecting reagents containing bleach. See page 4 for safety information.
- Perform all steps of the procedure at room temperature (15–25°C). During the procedure, work quickly.
- Perform all centrifugation steps at 20–25°C in a standard microcentrifuge. Ensure that the centrifuge does not cool below 20°C.
- In the procedure below, ▲ refers to use of starting volumes ≤100 µl, and ● refers to use of starting volumes of 100–200 µl.

Things to do before starting

- Buffer RPE is supplied as a concentrate. Before using for the first time, add 4 volumes of ethanol (96–100%) as indicated on the bottle to obtain a working solution.
- Buffer RLT may form a precipitate during storage. If necessary, redissolve by warming, and then place at room temperature (15–25°C).
- **Optional:** If cleaning up crude RNA preps (e.g., after salting-out methods) or samples rich in RNases, we recommend adding β -mercaptoethanol (β -ME) to Buffer RLT before use. Add 10 μ l β -ME per 1 ml Buffer RLT. Dispense in a fume hood and wear appropriate protective clothing. Buffer RLT containing β -ME can be stored at room temperature for up to 1 month. Alternatively, add 20 μ l of 2 M dithiothreitol (DTT) per 1 ml Buffer RLT. The stock solution of 2 M DTT in water should be prepared fresh or frozen in single-use aliquots. Buffer RLT containing DTT can be stored at room temperature for up to 1 month.

Procedure

1. Adjust the sample to a volume of ▲ 100 μ l or ● 200 μ l with RNase-free water. Add ▲ 350 μ l or ● 700 μ l Buffer RLT, and mix well.

If starting with an RNA pellet, be sure that the pellet is dissolved in the RNase-free water (supplied) before adding Buffer RLT.

Optional: Add β -ME (or DTT) to Buffer RLT before use (see “Things to do before starting”).

2. Add ▲ 250 μ l or ● 500 μ l of 96–100% ethanol to the diluted RNA, and mix well by pipetting. Do not centrifuge. Proceed immediately to step 3.
3. Transfer the sample (700 μ l) to an RNeasy MinElute spin column placed in a 2 ml collection tube (supplied). Close the lid gently, and centrifuge for 15 s at $\geq 8000 \times g$ ($\geq 10,000$ rpm). Discard the flow-through.*

For ● samples > 700 μ l, transfer the remaining sample (up to 700 μ l) and repeat the centrifugation. Discard the flow-through.*

* Flow-through contains Buffer RLT and is therefore not compatible with bleach. See page 4 for safety information.

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4. Place the RNeasy MinElute spin column in a new 2 ml collection tube (supplied). Add 500 μ l Buffer RPE to the spin column. Close the lid gently, and centrifuge for 15 s at $\geq 8000 \times g$ ($\geq 10,000$ rpm) to wash the spin column membrane. Discard the flow-through. Reuse the collection tube in step 5.

Note: Buffer RPE is supplied as a concentrate. Ensure that ethanol is added to Buffer RPE before use (see “Things to do before starting”).

5. Add 500 μ l of 80% ethanol to the RNeasy MinElute spin column. Close the lid gently, and centrifuge for 2 min at $\geq 8000 \times g$ ($\geq 10,000$ rpm) to wash the spin column membrane. Discard the flow-through and collection tube.

Note: After centrifugation, carefully remove the RNeasy MinElute spin column from the collection tube so that the column does not contact the flow-through. Otherwise, carryover of ethanol will occur.

6. Place the RNeasy MinElute spin column in a new 2 ml collection tube (supplied). Open the lid of the spin column, and centrifuge at full speed for 5 min. Discard the flow-through and collection tube.

To avoid damage to their lids, place the spin columns into the centrifuge with at least one empty position between columns. Orient the lids so that they point in a direction opposite to the rotation of the rotor (e.g., if the rotor rotates clockwise, orient the lids counterclockwise).

It is important to dry the spin column membrane since residual ethanol may interfere with downstream reactions. Centrifugation with the lids open ensures that no ethanol is carried over during RNA elution.

7. Place the RNeasy MinElute spin column in a new 1.5 ml collection tube (supplied). Add 14 μ l RNase-free water directly to the center of the spin column membrane. Close the lid gently, and centrifuge for 1 min at full speed to elute the RNA.

As little as 10 μ l RNase-free water can be used for elution if a higher RNA concentration is required, but the yield will be reduced by approximately 20%. Do not elute with less than 10 μ l RNase-free water as the spin column membrane will not be sufficiently hydrated.

The dead volume of the RNeasy MinElute spin column is 2 µl: elution with 14 µl RNase-free water results in a 12 µl eluate.

For RT-PCR and real-time RT-PCR with the purified RNA, QIAGEN offers a range of optimized, ready-to-use kits that provide highly specific and sensitive results. For details, visit www.qiagen.com/PCR. For whole transcriptome amplification (WTA) of limited amounts of RNA, we recommend the QuantiTect® Whole Transcriptome Kit. For details, visit www.qiagen.com/goto/WTA.

Protocol: Concentration of RNA Purified using the PAXgene Blood RNA Kit

This protocol is for the concentration of total cellular RNA purified using the PAXgene Blood RNA Kit (see ordering information, starting on page 34).

Determining the correct amount of starting material

A maximum of 45 µg RNA in a maximum volume of 200 µl can be concentrated in this protocol. This amount corresponds to the binding capacity of the RNeasy MinElute spin column. Before starting the procedure, RNA eluates obtained with the PAXgene Blood RNA Kit can be pooled as long as the total amount of RNA is ≤45 µg and the total volume is ≤200µl.

Important points before starting

- The RNA denaturation step (65°C incubation for 5 min) of the PAXgene Blood RNA Kit protocol does not need to be performed. RNA denaturation takes place in the last step of this protocol.
- Elution buffer BR5 (included in the PAXgene Blood RNA Kit) is required for RNA elution in this protocol.
- If preparing RNA for the first time, read Appendix A (page 21).
- DNase digestion does not need to be performed in this protocol because it has already been carried out in the PAXgene Blood RNA Kit protocol.
- Buffer RLT contains a guanidine salt and is therefore not compatible with disinfecting reagents containing bleach. See page 4 for safety information.
- Unless otherwise indicated, perform all steps of the procedure at room temperature (15–25°C). During the procedure, work quickly.
- Perform all centrifugation steps at 20–25°C in a standard microcentrifuge. Ensure that the centrifuge does not cool below 20°C.

- In the procedure below, ▲ refers to use of starting volumes $\leq 100 \mu\text{l}$, and ● refers to use of starting volumes of $100\text{--}200 \mu\text{l}$.

Things to do before starting

- Buffer RPE is supplied as a concentrate. Before using for the first time, add 4 volumes of ethanol (96–100%) as indicated on the bottle to obtain a working solution.
- Buffer RLT may form a precipitate during storage. If necessary, redissolve by warming, and then place at room temperature (15–25°C).

Procedure

1. Heat a heating block or water bath to 65°C for use in step 9.
2. Adjust the sample to a volume of ▲ 100 μl or ● 200 μl with RNase-free water. Add ▲ 350 μl or ● 700 μl Buffer RLT, and mix well.
3. Add ▲ 250 μl or ● 500 μl of 96–100% ethanol to the diluted RNA, and mix well by pipetting. Do not centrifuge. Proceed immediately to step 4.
4. Transfer the sample (700 μl) to an RNeasy MinElute spin column placed in a 2 ml collection tube (supplied). Close the lid gently, and centrifuge for 15 s at $\geq 8000 \times g$ ($\geq 10,000$ rpm). Discard the flow-through.*
For ● samples $>700 \mu\text{l}$, transfer the remaining sample (up to 700 μl) and repeat the centrifugation. Discard the flow-through.*
5. Place the RNeasy MinElute spin column in a new 2 ml collection tube (supplied). Add 500 μl Buffer RPE to the spin column. Close the lid gently, and centrifuge for 15 s at $\geq 8000 \times g$ ($\geq 10,000$ rpm) to wash the spin column membrane. Discard the flow-through.
Reuse the collection tube in step 6.

Note: Buffer RPE is supplied as a concentrate. Ensure that ethanol is added to Buffer RPE before use (see “Things to do before starting”).

* Flow-through contains Buffer RLT and is therefore not compatible with bleach. See page 4 for safety information.

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6. Add 500 μ l of 80% ethanol to the RNeasy MinElute spin column. Close the lid gently, and centrifuge for 2 min at $\geq 8000 \times g$ ($\geq 10,000$ rpm) to wash the spin column membrane. Discard the flow-through and collection tube.

Note: After centrifugation, carefully remove the RNeasy MinElute spin column from the collection tube so that the column does not contact the flow-through. Otherwise, carryover of ethanol will occur.

7. Place the RNeasy MinElute spin column in a new 2 ml collection tube (supplied). Open the lid of the spin column, and centrifuge at full speed for 5 min. Discard the flow-through and collection tube.

To avoid damage to their lids, place the spin columns into the centrifuge with at least one empty position between columns. Orient the lids so that they point in a direction opposite to the rotation of the rotor (e.g., if the rotor rotates clockwise, orient the lids counterclockwise).

It is important to dry the spin column membrane since residual ethanol may interfere with downstream reactions. Centrifugation with the lids open ensures that no ethanol is carried over during RNA elution.

8. Place the RNeasy MinElute spin column in a new 1.5 ml collection tube (supplied). Add 14 μ l of elution buffer BR5 (from the PAXgene Blood RNA Kit) directly to the center of the spin column membrane. Close the lid gently, and centrifuge for 1 min at full speed to elute the RNA.

As little as 10 μ l elution buffer BR5 can be used for elution if a higher RNA concentration is required, but the yield will be reduced by approximately 20%. Do not elute with less than 10 μ l elution buffer BR5 as the spin column membrane will not be sufficiently hydrated.

The dead volume of the RNeasy MinElute spin column is 2 μ l: elution with 14 μ l elution buffer BR5 results in a 12 μ l eluate.

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9. Incubate the eluate for 5 min at 65°C in a heating block or water bath. After incubation, chill immediately on ice.

Denaturation of the eluate is essential for maximum efficiency in downstream applications, such as RT-PCR, other amplification reactions or cDNA synthesis. It is not necessary to denature samples more than once, and samples remain denatured after freezing and thawing.

For RT-PCR and real-time RT-PCR with the purified RNA, QIAGEN offers a range of optimized, ready-to-use kits that provide highly specific and sensitive results. For details, visit www.qiagen.com/PCR. For whole transcriptome amplification (WTA) of limited amounts of RNA, we recommend the QuantiTect Whole Transcriptome Kit. For details, visit www.qiagen.com/goto/WTA.

Troubleshooting Guide

This troubleshooting guide may be helpful in solving any problems that may arise. For more information, see also the Frequently Asked Questions page at our Technical Support Center: www.qiagen.com/FAQ/FAQList.aspx. The scientists in QIAGEN Technical Services are always happy to answer any questions you may have about either the information and protocols in this handbook or sample and assay technologies (for contact information, see back cover or visit www.qiagen.com).

Comments and suggestions

Low or no recovery of RNA

- | | |
|---|--|
| a) RNase-free water (or elution buffer BR5) incorrectly dispensed | Pipet RNase-free water (or elution buffer BR5) to the center of the RNeasy MinElute spin column membrane to ensure that the membrane is completely covered. |
| b) Ethanol carryover | After the wash with 80% ethanol, be sure to centrifuge at full speed for 5 min to dry the RNeasy MinElute spin column membrane.

After centrifugation, carefully remove the RNeasy MinElute spin column from the collection tube so that the column does not contact the flow-through. Otherwise, carryover of ethanol will occur. |

Clogged RNeasy MinElute spin column

- | | |
|---------------------------------------|---|
| a) Too much starting RNA | In subsequent preps, reduce the amount of starting RNA. A maximum of 45 µg RNA can be used, including any carrier RNA. This amount corresponds to the binding capacity of an RNeasy MinElute spin column. |
| b) Centrifugation temperature too low | The centrifugation temperature should be 20–25°C. Some centrifuges may cool to below 20°C even when set at 20°C. This can cause formation of precipitates that can clog the spin column. If this happens, set the centrifugation temperature to 25°C. |

Comments and suggestions

Low A_{260}/A_{280} value

Water used to dilute RNA for A_{260}/A_{280} measurement

Use 10 mM Tris-Cl, pH 7.5, not RNase-free water, to dilute the sample before measuring purity (see Appendix B, page 24).

RNA degraded

RNase contamination

Although all RNeasy buffers have been tested and are guaranteed RNase-free, RNases can be introduced during use. Be certain not to introduce any RNases during RNA cleanup or later handling. See Appendix A (page 21) for general remarks on handling RNA.

Do not put RNA samples into a vacuum dryer or microcentrifuge that has been used in DNA preparations where RNases may have been used.

Adding β -ME to Buffer RLT may be helpful when cleaning up crude preps of RNA (e.g., after salting-out methods) or samples that contain large amounts of RNases (see protocols).

DNA contamination in downstream experiments

No DNase treatment

DNase digest the RNA sample before RNA cleanup (see Appendix C, page 28).

For real-time two-step RT-PCR experiments, carry out the RT step using the QuantiTect Reverse Transcription Kit, which provides cDNA synthesis with integrated removal of genomic DNA contamination. See ordering information, starting on page 34.

Comments and suggestions

RNA does not perform well in downstream experiments

Ethanol carryover

After the wash with 80% ethanol, be sure to centrifuge at full speed for 5 min to dry the RNeasy MinElute spin column membrane.

After centrifugation, carefully remove the RNeasy MinElute spin column from the collection tube so that the column does not contact the flow-through. Otherwise, carryover of ethanol will occur.

Appendix A: General Remarks on Handling RNA

Handling RNA

Ribonucleases (RNases) are very stable and active enzymes that generally do not require cofactors to function. Since RNases are difficult to inactivate and even minute amounts are sufficient to destroy RNA, do not use any plasticware or glassware without first eliminating possible RNase contamination. Great care should be taken to avoid inadvertently introducing RNases into the RNA sample during or after the purification procedure. To create and maintain an RNase-free environment, the following precautions must be taken during pretreatment and use of disposable and nondisposable vessels and solutions while working with RNA.

General handling

Proper microbiological, aseptic technique should always be used when working with RNA. Hands and dust particles may carry bacteria and molds and are the most common sources of RNase contamination. Always wear latex or vinyl gloves while handling reagents and RNA samples to prevent RNase contamination from the surface of the skin or from dusty laboratory equipment. Change gloves frequently and keep tubes closed whenever possible. Keep purified RNA on ice when aliquots are pipetted for downstream applications.

Disposable plasticware

The use of sterile, disposable polypropylene tubes is recommended throughout the procedure. These tubes are generally RNase-free and do not require pretreatment to inactivate RNases.

Nondisposable plasticware

Nondisposable plasticware should be treated before use to ensure that it is RNase-free. Plasticware should be thoroughly rinsed with 0.1 M NaOH, 1 mM EDTA* followed by RNase-free water (see “Solutions”, page 22). Alternatively, chloroform-resistant plasticware can be rinsed with chloroform* to inactivate RNases.

Glassware

Glassware should be treated before use to ensure that it is RNase-free. Glassware used for RNA work should be cleaned with a detergent,* thoroughly rinsed, and oven baked at 240°C for at least 4 hours (overnight, if more convenient) before use. Autoclaving alone will not fully inactivate many RNases. Alternatively, glassware can be treated with DEPC* (diethyl pyrocarbonate). Fill glassware with 0.1% DEPC (0.1% in water), allow to stand overnight (12 hours) at 37°C, and then autoclave or heat to 100°C for 15 minutes to eliminate residual DEPC.

Electrophoresis tanks

Electrophoresis tanks should be cleaned with detergent solution (e.g., 0.5% SDS),* thoroughly rinsed with RNase-free water, and then rinsed with ethanol† and allowed to dry.

Solutions

Solutions (water and other solutions) should be treated with 0.1% DEPC. DEPC is a strong, but not absolute, inhibitor of RNases. It is commonly used at a concentration of 0.1% to inactivate RNases on glass or plasticware or to create RNase-free solutions and water. DEPC inactivates RNases by covalent modification. Add 0.1 ml DEPC to 100 ml of the solution to be treated and shake vigorously to bring the DEPC into solution. Let the solution incubate for 12 hours at 37°C. Autoclave for 15 minutes to remove any trace of DEPC. DEPC will react with primary

* When working with chemicals, always wear a suitable lab coat, disposable gloves and protective goggles. For more information, consult the appropriate safety data sheets (SDSs), available from the product suppliers.

† Plastics used for some electrophoresis tanks are not resistant to ethanol. Take proper care and check the supplier's instructions.

amines and cannot be used directly to treat Tris* buffers. DEPC is highly unstable in the presence of Tris buffers and decomposes rapidly into ethanol and CO₂. When preparing Tris buffers, treat water with DEPC first, and then dissolve Tris to make the appropriate buffer. Trace amounts of DEPC will modify purine residues in RNA by carbethoxylation. Carbethoxylated RNA is translated with very low efficiency in cell-free systems. However, its ability to form DNA:RNA or RNA:RNA hybrids is not seriously affected unless a large fraction of the purine residues has been modified. Residual DEPC must always be eliminated from solutions or vessels by autoclaving or heating to 100°C for 15 minutes.

Note: RNeasy buffers are guaranteed RNase-free without using DEPC treatment and are therefore free of any DEPC contamination.

* When working with chemicals, always wear a suitable lab coat, disposable gloves and protective goggles. For more information, consult the appropriate safety data sheets (SDSs), available from the product suppliers.

Appendix B: Storage, Quantification and Determination of Quality of RNA

Storage of RNA

Purified RNA may be stored at -20°C or -70°C in RNase-free water. Under these conditions, no degradation of RNA is detectable after 1 year.

Quantification of RNA

The concentration of RNA can be determined by measuring the absorbance at 260 nm (A_{260}) in a spectrophotometer (see “Spectrophotometric quantification of RNA” below). For small amounts of RNA, however, it may not be possible to accurately determine amounts photometrically. Small amounts of RNA can be quantified using an Agilent® 2100 Bioanalyzer, fluorometric quantification or quantitative, real-time RT-PCR. When purifying RNA from particularly small samples (e.g., laser-microdissected samples), quantitative, real-time RT-PCR should be used for quantification.

Spectrophotometric quantification of RNA

To ensure significance, A_{260} readings should be greater than 0.15. An absorbance of 1 unit at 260 nm corresponds to 44 μg of RNA per ml ($A_{260} = 1 \rightarrow 44 \mu\text{g/ml}$). This relation is valid only for measurements at a neutral pH. Therefore, if it is necessary to dilute the RNA sample, this should be done in a buffer with neutral pH.* As discussed below (see “Purity of RNA”, page 25), the ratio between the absorbance values at 260 and 280 nm gives an estimate of RNA purity.

When measuring RNA samples, be certain that cuvettes are RNase-free, especially if the RNA is to be recovered after spectrophotometry. This can be accomplished by washing cuvettes

* When working with chemicals, always wear a suitable lab coat, disposable gloves and protective goggles. For more information, consult the appropriate safety data sheets (SDSs), available from the product supplier

with 0.1 M NaOH, 1 mM EDTA* followed by washing with RNase-free water (see “Solutions”, page 22). Use the buffer in which the RNA is diluted to zero the spectrophotometer. An example of the calculation involved in RNA quantification is shown below:

Volume of RNA sample = 10 μ l

Dilution = 1 μ l of RNA sample + 499 μ l of 10 mM Tris-Cl, * pH 7.0 (1/500 dilution)

Measure absorbance of diluted sample in a 1 ml cuvette (RNase-free)

$A_{260} = 0.2$

$$\begin{aligned}\text{Concentration of RNA sample} &= 44 \mu\text{g/ml} \times A_{260} \times \text{dilution factor} \\ &= 44 \mu\text{g/ml} \times 0.2 \times 500 \\ &= 4400 \mu\text{g/ml}.\end{aligned}$$

$$\begin{aligned}\text{Total amount} &= \text{concentration} \times \text{volume in milliliters} \\ &= 4400 \mu\text{g/ml} \times 0.01 \text{ ml} \\ &= 44 \mu\text{g of RNA}\end{aligned}$$

Purity of RNA

The ratio of the readings at 260 nm and 280 nm (A_{260}/A_{280}) provides an estimate of the purity of RNA with respect to contaminants that absorb in the UV spectrum, such as protein. However, the A_{260}/A_{280} ratio is influenced considerably by pH. Since water is not buffered, the pH and the resulting A_{260}/A_{280} ratio can vary greatly. Lower pH results in a lower A_{260}/A_{280} ratio and reduced sensitivity to protein contamination.[†] For accurate values, we recommend measuring absorbance in 10 mM Tris-Cl, pH 7.5. Pure RNA has an A_{260}/A_{280} ratio of 1.9–2.1[‡] in 10 mM Tris-Cl, pH 7.5. Always be sure to calibrate the spectrophotometer with the same solution used for dilution. For determination of RNA concentration, however, we recommend dilution of the sample in a buffer with neutral pH since the relationship between absorbance and

* When working with chemicals, always wear a suitable lab coat, disposable gloves and protective goggles. For more information, consult the appropriate safety data sheets (SDSs), available from the product suppliers.

[†] Wilfinger, W.W., Mackey, M., and Chomczynski, P. (1997) Effect of pH and ionic strength on the spectrophotometric assessment of nucleic acid purity. *BioTechniques* **22**, 474.

[‡] Values up to 2.3 are routinely obtained for pure RNA (in 10 mM Tris-Cl, pH 7.5) with some spectrophotometers.

concentration (A_{260} reading of 1 = 44 $\mu\text{g}/\text{ml}$ RNA) is based on an extinction coefficient calculated for RNA at neutral pH (see “Spectrophotometric quantification of RNA”, page 24).

DNA contamination

No currently available purification method can guarantee that RNA is completely free of DNA, even when it is not visible on an agarose gel. The level of genomic DNA contamination in an RNA sample depends on the method used for RNA purification. For example, RNA purified by phenol-chloroform extraction and alcohol precipitation will contain small amounts of DNA, while RNA purified using the PAXgene Blood RNA Kit (which includes on-column DNase digestion) will contain negligible amounts of DNA.

When analyzing low-abundance transcripts by real-time RT-PCR, any interference by residual DNA contamination can be detected by performing a control reaction in which reverse transcriptase is omitted in the RT step. To prevent any interference by DNA contamination in real-time RT-PCR, we recommend designing primers that anneal at an exon–exon boundary so that genomic DNA will not be amplified. QuantiTect Primer Assays from QIAGEN are designed for SYBR® Green-based real-time RT-PCR analysis of RNA sequences (without detection of genomic DNA) where possible (the assays can be ordered online at www.qiagen.com/GeneGlobe).

For real-time RT-PCR assays where amplification of genomic DNA cannot be avoided, and for other experiments where RNA must be free of genomic DNA contamination, we recommend treating RNA samples with DNase. If handling RNA redissolved in water or buffer, we recommend performing a DNase digestion in solution (see Appendix C, page 28) before starting the RNA cleanup procedure on page 10. However, if the RNA was isolated by phenol-chloroform extraction, the aqueous phase containing RNA should be subjected to RNA cleanup with on-column DNase digestion using the RNeasy Micro Kit (cat. no. 74004).

As an alternative to DNase digestion, the QuantiTect Reverse Transcription Kit provides fast cDNA synthesis with integrated removal of genomic DNA contamination (see ordering

information, starting on page 34). The kit is dedicated for use in real-time two-step RT-PCR, providing high yields of cDNA representing all transcript regions.

Integrity of RNA

The integrity and size distribution of total RNA purified with RNeasy Kits can be checked by denaturing agarose gel electrophoresis and ethidium bromide staining* or by using an Agilent 2100 Bioanalyzer. The respective ribosomal RNAs should appear as sharp bands or peaks. The apparent ratio of 28S rRNA to 18S rRNA should be approximately 2:1. If the ribosomal bands or peaks of a specific sample are not sharp, but appear as a smear towards smaller sized RNAs, it is likely that the RNA sample suffered major degradation either before or during RNA purification.

* When working with chemicals, always wear a suitable lab coat, disposable gloves and protective goggles. For more information, consult the appropriate safety data sheets (SDSs), available from the product supplier.

Appendix C: DNase Digestion of RNA before RNA Cleanup

This protocol describes how to digest contaminating DNA in RNA solutions prior to RNA cleanup and concentration. This protocol requires use of the QIAGEN RNase-Free DNase Set (see ordering information, starting on page 34; see the RNase-Free DNase Set product insert for product description and more information).

Important points before starting

- Do not vortex reconstituted DNase I. DNase I is especially sensitive to physical denaturation. Mixing should only be carried out by gently inverting the vial.

Things to do before starting

- Prepare DNase I stock solution before using the RNase-Free DNase Set for the first time. Dissolve the lyophilized DNase I (1500 Kunitz units) in 550 μ l of the RNase-free water provided. To avoid loss of DNase I, do not open the vial. Inject RNase-free water into the vial using an RNase-free needle and syringe. Mix gently by inverting the vial. Do not vortex.
- For long-term storage of reconstituted DNase I, remove the stock solution from the glass vial, divide it into single-use aliquots, and store at -20°C for up to 9 months. Thawed aliquots can be stored at $2-8^{\circ}\text{C}$ for up to 6 weeks. Do not refreeze the aliquots after thawing.

Procedure

C1. Mix the following in a microcentrifuge tube:

- ≤87.5 µl RNA solution (contaminated with genomic DNA)
- 10 µl Buffer RDD
- 2.5 µl DNase I stock solution

Make the volume up to 100 µl with RNase-free water.

The reaction volumes can be doubled if necessary (to 200 µl final volume).

C2. Incubate on the benchtop (20–25°C) for 10 min.

C3. Clean up the RNA according to “Protocol: RNA Cleanup and Concentration” on page 10.

Appendix D: RNA Cleanup after Lysis and Homogenization with QIAzol[®] Lysis Reagent

QIAzol Lysis Reagent is a monophasic solution of phenol and guanidine thiocyanate that can be used for sample lysis and partial purification of total RNA (see ordering information, starting on page 34). Cleanup using the RNeasy MinElute Cleanup Kit is recommended to remove any contaminating phenol. The aqueous phase from QIAzol lysis is used as starting material in this protocol. It is therefore not necessary to precipitate the RNA and redissolve it prior to RNA cleanup.*

Determining the correct amount of starting material

This protocol is designed for QIAzol preps with a maximum starting volume of 1 ml QIAzol Lysis Reagent. This corresponds to a final volume of approximately 600 μ l (aqueous phase) for RNA cleanup.

A maximum of 45 μ g RNA can be cleaned up in this protocol. This amount corresponds to the binding capacity of the RNeasy MinElute spin column. If the expected RNA yield is >45 μ g, use an appropriate proportion of the QIAzol lysate per RNeasy MinElute spin column.

* This protocol also works well with some other reagents containing phenol and guanidine thiocyanate. Please contact QIAGEN Technical Services for more details (see back cover for contact information).

Important points before starting

- If preparing RNA for the first time, read Appendix A (page 21).
- Generally, DNase digestion is not required since the combination of QIAzol and RNeasy MinElute technologies efficiently removes most of the DNA without DNase treatment. However, further DNA removal may be necessary for certain RNA applications that are sensitive to very small amounts of DNA (e.g., TaqMan RT-PCR analysis with a low-abundance target). If DNase digestion is required, we recommend performing RNA cleanup with the RNeasy Micro Kit (cat. no. 74004), which is supplied with the RNase-Free DNase Set and Buffer RW1 for on-column DNase digestion. Please note that the combination of the RNeasy MinElute Cleanup Kit and the RNase-Free DNase Set cannot be used for RNA cleanup with on-column DNase digestion as they are not supplied with Buffer RW1.
- QIAzol Lysis Reagent contains a guanidine salt and is therefore not compatible with disinfecting reagents containing bleach. See the *QIAzol Handbook* for safety information.
- Unless otherwise indicated, perform all steps of the procedure at room temperature (15–25°C). During the procedure, work quickly.
- Perform all centrifugation steps at 20–25°C in a standard microcentrifuge. Ensure that the centrifuge does not cool below 20°C.

Things to do before starting

- Buffer RPE is supplied as a concentrate. Before using for the first time, add 4 volumes of ethanol (96–100%) as indicated on the bottle to obtain a working solution.

Procedure

- D1. Carry out homogenization of the sample in QIAzol Lysis Reagent, followed by phase separation, as described in the *QIAzol Handbook* (steps 1–7 of the QIAzol protocol for lysis and homogenization).

D2. Transfer the upper, aqueous phase to a new collection tube. Add 1 volume of 70% ethanol, and mix thoroughly by vortexing. Do not centrifuge. Proceed immediately to step D3.

D3. Transfer up to 700 μ l of the sample to an RNeasy MinElute spin column placed in a 2 ml collection tube (supplied). Close the lid gently, and centrifuge for 15 s at $\geq 8000 \times g$ ($\geq 10,000$ rpm). Discard the flow-through.*

If the sample is $>700 \mu$ l, transfer the remaining sample (up to 700 μ l) and repeat the centrifugation. Discard the flow-through.*

D4. Place the RNeasy MinElute spin column in a new 2 ml collection tube (supplied). Add 500 μ l Buffer RPE to the spin column. Close the lid gently, and centrifuge for 15 s at $\geq 8000 \times g$ ($\geq 10,000$ rpm) to wash the spin column membrane. Discard the flow-through. Reuse the collection tube in step D5.

Note: Buffer RPE is supplied as a concentrate. Ensure that ethanol is added to Buffer RPE before use (see “Things to do before starting”).

D5. Add 500 μ l of 80% ethanol to the RNeasy MinElute spin column. Close the lid gently, and centrifuge for 2 min at $\geq 8000 \times g$ ($\geq 10,000$ rpm) to wash the spin column membrane. Discard the flow-through and collection tube.

Note: After centrifugation, carefully remove the RNeasy MinElute spin column from the collection tube so that the column does not contact the flow-through. Otherwise, carryover of ethanol will occur.

* Flow-through contains QIAzol Lysis Reagent and is therefore not compatible with bleach. See the *QIAzol Handbook* for safety information.

D6. Place the RNeasy MinElute spin column in a new 2 ml collection tube (supplied). Open the lid of the spin column, and centrifuge at full speed for 5 min. Discard the flow-through and collection tube.

To avoid damage to their lids, place the spin columns into the centrifuge with at least one empty position between columns. Orient the lids so that they point in a direction opposite to the rotation of the rotor (e.g., if the rotor rotates clockwise, orient the lids counterclockwise).

It is important to dry the spin column membrane since residual ethanol may interfere with downstream reactions. Centrifugation with the lids open ensures that no ethanol is carried over during RNA elution.

D7. Place the RNeasy MinElute spin column in a new 1.5 ml collection tube (supplied). Add 14 μ l RNase-free water directly to the center of the spin column membrane. Close the lid gently, and centrifuge for 1 min at full speed to elute the RNA.

As little as 10 μ l RNase-free water can be used for elution if a higher RNA concentration is required, but the yield will be reduced by approximately 20%. Do not elute with less than 10 μ l RNase-free water as the spin column membrane will not be sufficiently hydrated.

The dead volume of the RNeasy MinElute spin column is 2 μ l: elution with 14 μ l RNase-free water results in a 12 μ l eluate.

For RT-PCR and real-time RT-PCR with the purified RNA, QIAGEN offers a range of optimized, ready-to-use kits that provide highly specific and sensitive results.

For details, visit www.qiagen.com/PCR. For whole transcriptome amplification (WTA) of limited amounts of RNA, we recommend the QuantiTect Whole Transcriptome Kit. For details, visit www.qiagen.com/goto/WTA.

Ordering Information

Product	Contents	Cat. no.
RNeasy MinElute Cleanup Kit (50)	50 RNeasy MinElute Spin Columns, Collection Tubes, RNase-Free Reagents and Buffers	74204
Accessories		
RNase-Free DNase Set (50)	1500 units RNase-Free DNase I, RNase-Free Buffer RDD and RNase-Free Water	79254
QIAcube Connect — for fully automated nucleic acid extraction with QIAGEN spin-column kits		
QIAcube Connect*	Instrument, connectivity package, 1-year warranty on parts and labor	Inquire
Starter Pack, QIAcube	Filter-tips, 200 µl (1024), 1000 µl filter-tips (1024), 30 ml reagent bottles (12), rotor adapters (240), elution tubes (240), rotor adapter holder	990395
Related products		
RNeasy Plus Micro Kit — for purification of total RNA from small cell and tissue samples using gDNA Eliminator columns		
RNeasy Plus Micro Kit (50)	50 RNeasy MinElute Spin Columns, 50 gDNA Eliminator Mini Spin Columns, Collection Tubes, Carrier RNA, RNase-Free Reagents and Buffers	74034

RNeasy Mini Kit — for purification of total RNA from cells, tissues and yeast and for RNA cleanup

RNeasy Mini Kit (50) [†]	50 RNeasy Mini Spin Columns, Collection Tubes, RNase-Free Reagents and Buffers	74104
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PAXgene Blood RNA Kit — for isolation and purification of intracellular RNA from whole blood stabilized in PAXgene Blood RNA Tubes

PAXgene Blood RNA Kit (50)	50 PAXgene Spin Columns, 50 PAXgene Shredder Spin Columns, Processing Tubes, RNase-Free DNase Set, RNase-Free Reagents and Buffers. To be used in conjunction with PAXgene Blood RNA Tubes [‡]	762164 [§] 762174 [¶]
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QIAzol Lysis Reagent — for efficient lysis of fatty tissues and all other types of tissue before RNA purification

QIAzol Lysis Reagent (200 ml)	200 ml QIAzol Lysis Reagent	79306
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QuantiTect Reverse Transcription Kit — for fast reverse transcription for sensitive real-time two-step RT-PCR

QuantiTect Reverse Transcription Kit (50) [†]	For 50 x 20 µl reactions: gDNA Wipeout Buffer, Quantiscript [®] Reverse Transcriptase, Quantiscript RT Buffer, RT Primer Mix and RNase-Free Water	205311
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QuantiTect Primer Assays — for use in real-time RT-PCR with SYBR Green detection (search for and order assays at www.qiagen.com/GeneGlobe)

QuantiTect Primer Assay (200)	For 200 x 50 µl reactions or 400 x 25 µl reactions: 10x QuantiTect Primer Assay (lyophilized)	Varies
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QuantiFast[®] SYBR Green PCR Kit — for fast, quantitative, real-time PCR and two-step RT-PCR using SYBR Green I

QuantiFast SYBR Green PCR Kit (400) [†]	For 400 x 25 µl reactions: 3 x 1.7 ml 2x Master Mix (contains ROX dye), 2 x 2 ml RNase-Free Water	204054
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Quantifast SYBR Green RT-PCR Kit — for fast, quantitative, real-time one-step RT-PCR using SYBR Green I

Quantifast SYBR Green RT-PCR Kit (400) [†]	For 400 x 25 µl reactions: 3 x 1.7 ml 2x Master Mix (contains ROX dye), 100 µl RT Mix, 2 x 2 ml RNase-Free Water	204154
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Quantifast Probe PCR Kits — for fast, quantitative, real-time PCR and two-step RT-PCR using sequence-specific probes

For all instruments from Applied Biosystems except the Applied Biosystems® 7500:

Quantifast Probe PCR Kit (400) [†]	For 400 x 25 µl reactions: 3 x 1.7 ml 2x Master Mix (contains ROX dye), 2 x 2 ml RNase-Free Water	204254
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For the Applied Biosystems 7500 and instruments from Bio-Rad/MJ Research, Cepheid, Corbett Research, Eppendorf, Roche and Stratagene:

Quantifast Probe PCR +ROX Vial Kit (400) [†]	For 400 x 25 µl reactions: 3 x 1.7 ml 2x Master Mix (without ROX dye), 210 µl ROX Dye Solution, 2 x 2 ml RNase-Free Water	204354
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Quantifast Probe RT-PCR Kits — for fast, quantitative, real-time one-step RT-PCR using sequence-specific probes

For all instruments from Applied Biosystems except the Applied Biosystems 7500:

Quantifast Probe RT-PCR Kit (400) [†]	For 400 x 25 µl reactions: 3 x 1.7 ml 2x Master Mix (contains ROX dye), 100 µl RT Mix, 2 x 2 ml RNase-Free Water	204454
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For the Applied Biosystems 7500 and instruments from Bio-Rad/MJ Research, Cepheid, Corbett Research, Eppendorf, Roche and Stratagene:

Quantifast Probe RT-PCR +ROX Vial Kit (400) [†]	For 400 x 25 µl reactions: 3 x 1.7 ml 2x Master Mix (without ROX dye), 210µl ROX Dye Solution, 100 µl RT Mix, 2 ml RNase-Free Water	204554
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* All QIAcube Connect instruments are provided with a region-specific connectivity package, including tablet and equipment necessary to connect to the local network. Further, QIAGEN offers comprehensive instrument service products, including service agreements, installation, introductory training and preventive subscription. Contact your local sales representative to learn about your options.

† Larger kit size and format available; for details, visit www.qiagen.com/.

‡ PAXgene Blood RNA Tubes (cat. no. 762165) are available from BD and BD authorized distributors (www.bd.com).

§ Canada and USA.

¶ Rest of the world; kit not available in all countries.

For up-to-date licensing information and product-specific disclaimers, see the respective QIAGEN kit handbook or user manual. QIAGEN kit handbooks and user manuals are available at www.qiagen.com or can be requested from QIAGEN Technical Services or your local distributor.

Visit www.qiagen.com/geneXpression to find out more about standardized solutions for gene expression analysis — from RNA preparation to real-time RT-PCR

Document Revision History

Date	Changes
January 2020	Updated text, ordering information and intended use for QIAcube Connect.

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QIAzol Lysis Reagent is a subject of US Patent No. 5,346,994 and foreign equivalents.

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Notes

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